

The plant growth regulator methyl jasmonate inhibits aflatoxin B₁ production by *Aspergillus parasiticus* in caper

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To cite this article:

Dido Maria Meimaroglou, Dia Galanopoulou, Fotini Flouri, Panagiota Markaki. The Plant Growth Regulator Methyl Jasmonate Inhibits Aflatoxin B₁ Production by *Aspergillus Parasiticus* in Caper. *International Journal of Nutrition and Food Sciences*. Special Issue: Food Safety. Vol. 3, No. 5-1, 2014, pp. 10-17. doi: 10.11648/j.ijjnfs.s.2014030501.13

Abstract: Aflatoxins, produced by some aflatoxigenic strains of the *Aspergillus* species, are known as potent carcinogenic. Aflatoxin biosynthesis involves lipid peroxidation with the presence of fatty acid hydroperoxides promoting aflatoxin production. Methyl jasmonate (MeJA) derives from α -linolenic acid and is a plant growth regulator, produced as a response to stress, such as by environment or pathogen attack. This study reports on the effect of MeJA added on *A. parasiticus* growth and AFB₁ production in caper, an edible plant of Greek origin used as condiment. AFB₁ determination in caper was performed by using HPLC-FD. Five different concentrations of MeJA, 10⁻⁶ M, 10⁻⁴ M, 10⁻³ M, 10⁻² M, and 10⁻¹ M, were added in caper samples and the kinetic of the AFB₁ production by *A. parasiticus* was studied for an incubation period of 15 days. Results revealed that MeJA affects AFB₁ production by *A. parasiticus* in a dose-dependent manner. MeJA at a concentration of 10⁻⁶ M stimulated AFB₁ production after the 9th day of incubation. MeJA at concentrations of 10⁻⁴ M and 10⁻³ M decreased moderately AFB₁ output. Finally, MeJA added to the caper samples at a concentration of 10⁻² M and 10⁻¹ M inhibited AFB₁ by 97.74% and 98.42% respectively on the 12th day of observation.

Keywords: Aflatoxin B₁, Methyl Jasmonate, *Aspergillus Parasiticus*, Caper, HPLC

1. Introduction

Aflatoxins are polyketide secondary fungal metabolites produced by the toxigenic strains of *Aspergillus flavus*, *Aspergillus parasiticus* and *Aspergillus nomius* and they are known as potent carcinogenic, teratogenic as well as genotoxic mycotoxins [1, 2]. Among the four naturally occurring aflatoxins, Aflatoxin B₁ (AFB₁) is the most potent one [3].

Aflatoxin contamination of food and feeds infected with *Aspergillus* spp. poses a worldwide food safety problem [4]. Seeds in particular, are susceptible and the lipid content of seeds plays a part in determining the severity of aflatoxin contamination [e.g. 5]. Recent reports support an aflatoxin/lipid relationship. It was shown by [6] that cottonseed meal sustained 1000-fold less aflatoxin production by *A. flavus* after lipid extraction.

The plant hormones jasmonic acid and methyl jasmonate,

along with their intermediate compounds, produced via the octadecanoid pathway, are important signaling molecules called jasmonates. These are widespread in the plant kingdom and play crucial roles in biotic/abiotic stress responses, as well as in processes related to plant growth and development. In plants, jasmonates are synthesized as a response to systemic or localized signals similar to oligosaccharides released from fungi or plant cell walls during plant-pathogen interactions [7]. According to [8], phytopathogenic bacterial species have developed specific methods in order to attack plant cells and to use plant substances for their own growth. [9] have reported that jasmonic acid levels increased rapidly in response to biotic and abiotic stress such as mechanical stress. Lately, natural elicitors were combined with methyl jasmonate (MeJA) to evaluate its effects on phytoalexin and AFB₁ production in cotton plants [10]. According to [11], the levels of endogenous hormones decreased in seeds of *Zea mays* treated with AFB₁, while [12] reported that substances such as aflatoxins inhibit

the growth and development process in plants.

As far as *Aspergilli* are concerned, interactions between jasmonates, *Aspergillus* growth and aflatoxin production are likely. These interactions are of great interest as they suggest that there is a mechanism involving plant LOX pathways that affects aflatoxin biosynthesis. Once the fungus has invaded the seed, it first destroys the lipid bodies, which are primarily composed of palmitic, oleic, and linolenic acid [13, 14]. Many in vitro studies have shown that lipid oxidation affects aflatoxin biosynthesis [5, 15, 16]. Linoleic as well as linolenic acid can undergo a regio- and stereospecific oxygenation catalyzed by the widely distributed plant stress response enzyme lipoxygenase (LOX) [17] to yield 13S-hydroperoxy-cis-9-trans-11-octadecadienoic acid (13S-HPODE) and 13S-hydroperoxycis-9, trans-11, cis-15-octadecatrienoic acid (13S-HPOTE), respectively [18]. These fatty acid hydroperoxides are further converted through the octadecanoid pathway to jasmonates [19].

Capparis sicula Veill. in Duham, *C. orientalis* Veill. in Duham and *C. spinosa* L. are small shrubs widely distributed in Mediterranean countries. Their beneficial properties, including antirheumatic, tonic as well as expectorant activities, have been known since antiquity from numerous populations of different countries around the Mediterranean Sea [20]. The flower buds and the fruits of *C. spinosa* and *C. sicula* have been used as a condiment in Greece since ancient times. Capers are usually commercialized in some Mediterranean countries and they are exported mainly to central European countries, the USA and the UK as a delicatessen product. From a dietary point of view, the flavonoid content and the antioxidant (free radical scavenging) activity of these compounds give a special value to this product [21]. A great deal of attention has been given to *Capparis* spp. buds, fruits, young shoots and seed oil. [22] reported on caper seeds growing in Turkey with oleic and linoleic acid being the major unsaturated fatty acids, while palmitic acid was the predominant saturated one.

With an oil content of approximately 30%, seeds of *C. ovata* and *C. spinosa* could be interesting sources for the production of vegetable oil. The results indicate that the oil contains linoleic acid as the major fatty acid accompanied by oleic acid and its isomer vaccenic acid. The content and composition of tocopherols are comparable to those of other sources such as rapeseed and sunflower oil. The relatively high content of Δ^5 -avenasterol is interesting, as it has been suggested as an antioxidant and antipolymerization agent in frying oils [23].

In literature there is no information concerning the AFB₁ production in capers, a plant with a high lipid content, as mentioned above. Therefore, interactions between MeJA concentrations and AFB₁ production, using a natural substrate such as capers inoculated with *A. parasiticus*, are investigated in the present work. More precisely it was examined which MeJA concentrations inhibit the AFB₁ production. Additionally, in the present work the development and validation of a method determining AFB₁ in capers is evaluated.

2. Materials and Methods

2.1. Apparatus

A laminar flow (Telstar Bio IIA, Madrid, Spain); an autoclave, Selecta Autester-E Dry (PBI Milano, Italy), an incubator WTB Binder (Tuttingen, Germany) and a centrifuge Sorvall RC-5B (HS-4) (Norwalk, USA) were used during this study. HPLC was performed using a Hewlett-Packard 1050 (Waldborn, Germany) liquid chromatograph equipped with a JASCO FP-920 (Japan) fluorescence detector and an HP integrator 3395. The HPLC column used was a C18 Nova-Pak (60 Å, 4 µm, 4.6 x 250 mm).

The mobile phase for AFB₁ determination [water + acetonitrile + methanol (20+4+3)] was filtered through Millipore HVLP filters (0.45 µm) before use. Detection of the AFB₁ hemiacetal derivative (AFB_{2a}) was carried out at $\lambda_{\text{ex}} = 365$ nm and $\lambda_{\text{em}} = 425$ nm. The flow rate was 1 ml min⁻¹ while the retention time was 12.60 (±0.2) min for AFB₁ in caper.

2.2. Reagents

The AFB₁ standard was purchased from Sigma-Aldrich (St. Louis, MO, USA). The filters and the C₁₈ Nova-Pak HPLC column were from Waters (Millipore, Milford, MA, USA). The Aflatest immunoaffinity columns were from Vicam and HPLC solvents were of HPLC grade (LABSCAN, Dublin, Ireland). Trifluoroacetic acid was purchased from Merck (Darmstadt, Germany). The purity of MeJA used was tested by GC analysis using a Hewlett-Packard GC (flame ionization detector) on a BPX70-coated fused-silica capillary column [24].

2.3. Media

Aspergillus flavus parasiticus agar (AFPA) was prepared by dissolving 4 g of yeast extract (Oxoid) (Basingstoke, Hampshire, England), 2 g of bacteriological peptone (Oxoid), 0.1 g of ferric ammonium citrate, 0.2 ml of Dichloran (0.2% in ethanol, Fluka, Neu-Ulm, Switzerland), 0.2 g of chloramphenicol (Oxoid) and 3 g agar (Oxoid) in 200 ml of distilled water, final pH 0.0-0.5 [25]. Czapek Dox agar (CZA) was prepared by dissolving 0.4 g of sodium nitrite, 0.1 g of potassium chloride, 0.1 g of magnesium sulfate, 0.002 g of ferric sulfate, 0.2 g of dipotassium phosphate, 6 g of sucrose, 3 g of agar, 0.002 g of zinc sulfate, and 0.001 g of copper sulfate in 200 ml of distilled water, final pH 6.0-6.5 [26].

2.4. Preparation of Spore Inoculum

The aflatoxigenic strain *A. parasiticus* sp (IMI 283883) utilized throughout this study was obtained from the International Mycological Institute (Engham, Surrey, UK). An inoculum was obtained by growing the mold on a slant of stock cultures of CZA, which were maintained at 5°C [27,28]. Spore inoculum was prepared by growing *A. parasiticus* on CZA for 7 days at 30°C and spores were harvested aseptically using 10 ml of sterile 0.01 % v/v Tween 80 solution [29]. AFB₁ transfer from the initial culture was minimized by

centrifuging the spore suspension (1000 g for 1 min) and resuspending the spore in 10 ml of sterile Tween 80 solution twice. Dilutions (10^{-1} , 10^{-2} , 10^{-3} , and 10^{-4}) from the initial spore suspension were prepared in sterile tubes containing 10 ml of 0.05% Tween 80 (v/v) suspension. The spore concentration was determined by the spread plate surface count technique, using 0.1 ml of each dilution on four AFPA plates [25, 30] after incubation at 30° C for 2 days. The population size was estimated by the reverse intense yellow/orange coloration of the colonies. In order to obtain an inoculum containing 10^2 conidia, plates with 10-100 colony forming units (cfu) were selected and the desired 10^2 spore quantity used in the present study was estimated. The quantity of 10^2 spores / flask was chosen as it was the minimum concentration found in the literature producing detectable amounts of AFB₁ by *Aspergillus* [31].

2.5. Inoculation

Three flasks, containing 15 g of capers, were used for each case and for each day of observation as shown in Table 1. MeJA in ethanol at final concentrations of 10^{-1} , 10^{-2} , 10^{-3} , 10^{-4} and 10^{-6} M flask⁻¹ was added into each of the three flasks containing caper (non inoculated) for each day of observation. All flasks, including control samples (both inoculated or not) as well as samples both treated with MeJA or not and samples inoculated with *A. parasiticus* and non-inoculated, were incubated under stationary conditions at 30° C. Immediately after autoclaving for 30 minutes at 115° C, as it is suggested for safety reasons [32], AFB₁ was determined on days 0, 3, 7, 9, 12 and 15 of incubation. The experiment was repeated in triplicate

Table 1. Inoculated samples of caper

Samples of caper (15 g) inoculated with <i>A.parasiticus</i> with MEJA addition at different concentrations
Caper without MeJA addition , non- inoculated + ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + (10^{-6} M) corresponding to 0.0022mg MEJA in ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + (10^{-4} M) corresponding to 0.227mg MEJA in ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + (10^{-3} M) corresponding to 2.267mg MEJA in ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + (10^{-2} M) corresponding to 22.667 mg MEJA in ethanol
Caper + 100 conidia of <i>A.parasiticus</i> + (10^{-1} M) corresponding to 226.667mg MEJA in ethanol

Three flasks were used for each case and for each day of observation. All samples were incubated during 15 days and AFB₁ was determined on days 0, 3, 7, 9, 12, 15.

2.6. Sampling and Treatment

An initial amount of 10 kilograms of capers originated from Cyclades, which are the most popular in the market, were purchased from the market of Syros in the summer of 2011.

The capers were washed with water, cut into pieces and, in order to eliminate their natural microbiota, they were treated

before inoculation with *A. parasiticus* as follows: dipped in 70% ethanol and shaken for 1 min. Following that, ethanol is rejected. Then washed with 1.25% NaOCl for three minutes and rinsed with sterile distilled water twice. After treatment, representative samples of 15 gr from the damaged capers (in order to obtain samples with similar natural microbiota) were aseptically employed into sterile flasks forming a solid mass [33].

2.7. AFB₁ Determination Derivatization and HPLC Analysis

The content of each flask containing caper was mixed with 30 ml of methanol water (80-20) and well-shaken for 10 min. After filtration, an aliquot of 1 ml from each flask was used for AFB₁ analysis. The 1 ml-aliquot from the filtrate was mixed with 10 ml of distilled water. The mixture was transferred onto an Aflatest immunoaffinity column (flow rate 6 ml min⁻¹) and washed twice with 10 ml of distilled water. The column was then allowed to dry by passing air through it. AFB₁ was eluted with 2 ml of acetonitrile (flow rate: 0.3 ml min⁻¹). Before derivatization, the eluate was evaporated to dryness on a water bath under a gentle steam of nitrogen [34].

A derivative of AFB₁ (AFB_{2a}, hemiacetal of AFB₁) was prepared by adding 200 µl of hexane and 200 µl of trifluoroacetic acid to the evaporated solution of AFB₁ eluate, heating for 10 min at 40° C in a water bath, evaporating to dryness under nitrogen, redissolving in an appropriate volume of water-acetonitrile (9:1) to give a concentration of <10 ng ml⁻¹ and analyzing by HPLC (volume injected: 40 µl). AFB_{2a} shows enhanced fluorescence compared to AFB₁ [35]. In addition to that, AFB_{2a} is less toxic compared to AFB₁ because of its protein binding properties [36].

2.8. Statistical Analysis

Data were analyzed by one-way ANOVA and t-test. The mean differences, which are significantly different, were examined by using the Tukey's test [37].

3. Results and Discussion

3.1. Characterization of the AFB₁ Determination in Capers

In the present study, the analytical protocol for the determination of AFB₁ in capers was in-house characterized regarding the following criteria: linearity, accuracy, repeatability, internal reproducibility, limits of detection (DL) and quantification (QL). Additionally, the limits of repeatability (r) and reproducibility(R) were calculated as following: $r = 2.8 \times SDr$ and $R = 2.8 \times SDR$, respectively. As far as the limits of detection (DL) and quantification (QL) are concerned, they were calculated using two different ways: (1) according to the following formulas $DL = [b_0 + 3S(b_0)]/b_1$, $QL = [b_0 + 10S(b_0)]/b_1$, where b_0 is the response of the blank (intercept of the calibration model), $S(b_0)$ is the standard deviation of the blank and b_1 is the sensitivity (calibration model slope) and (2) as the signal-to-noise ratio 3:1. The accuracy was estimated by analyzing samples (15 g) of caper

spiked with different quantities of AFB₁. Furthermore, a Fisher test was applied to confirm the acceptability of the linear regression. If the F1 ratio is greater than the critical

value F [1, p (n- 1), 1- a] corresponding to a Fisher variable with a risk a = 0.1% for 1 and p (n - 1) degrees of freedom, the regression model can be considered as acceptable.

Table 2. Accuracy, recovery, detection limit of the AFB₁ determination in caper

Samples	AFB ₁ (ng g ⁻¹) added					Recovery of the method%	RSD%	Detection Limit ng g ⁻¹ caper
	2	5	10	15	20			
AFB ₁ (ng g ⁻¹) recovered						79.00	6.53	0.06
1	1.82	4.45	8.14	12.91	16.90			
2	1.87	4.48	7.90	12.98	14.96			
3	1.88	4.50	7.75	12.47	16.70			
4	1.81	4.52	8.25	12.24	15.44			
Average	1.85	4.49	8.01	12.65	16.00	y = 0.36 (±0.27) + 0.79 (±0.02) x R = 0.9984		
SD	0.04	0.03	0.23	0.35	0.95			
RSD%	1.90	0.66	2.83	2.80	5.92			

Thereupon, the hypothesis must be checked if the lack of fit of the model is negligible and that it is a straight line throughout the chosen field. If the F1 ratio is less or equal to the critical value F[p - 2, p (n- 1), 1 - a] corresponding to a Fisher variable with a risk of 0.1% for p - 2 and p (n - 1) degrees of freedom, the field of linearity chosen can then be approved. For the determination, p = 5 levels and n = 4 repetitions were used (Table 2). Regarding the linearity, an AFB₁ standard solution (5 µg ml⁻¹ in methanol) was prepared by diluting 10 µl from an AFB₁ stock solution (0.5 mg ml⁻¹ in methanol) measuring its absorbance at 335 nm (A = 0.385). Thereupon, solutions of AFB₁ in mobile phase were prepared at concentrations of 10, 5, 2.5, 2 and 1 ng ml⁻¹. The volume injected into the column was 40 µl. Linear regression was used to prepare standard curves by using the peak areas mean values of five injections for each one of the five solutions. The equation of the standard curve was $y = -2.57(\pm 0.69) 10^5 + 9.6(\pm 0.13) 10^5 x$, where y is the peak area of AFB₁ injected (40 µl) and x is the amount in ng of standard AFB₁ injected (40 µl).

As far as the repeatability is concerned, this parameter was estimated by analyzing five subsamples (15 g) of capers spiked with 100 ng of AFB₁ under repetitive conditions (mean recovered = 97.36, RSD = 6.31%). The internal reproducibility was also estimated by determining five samples (15 g) of capers at time intervals (at five different days of the month). The samples were spiked with 100 ng AFB₁ (mean recovered = 95.54, RSD = 5.76%). In the present study the limits of repeatability (r) and reproducibility (R) were found to be: r = 17.19 and R = 15.40, respectively (SDr = 6.14 and SDR = 5.50).

The accuracy of the method applied in capers was studied by analyzing samples (15 g) of capers spiked with AFB₁ at different concentrations corresponding to AFB₁ ng g⁻¹ being shown in Table 2.

The regression coefficient r of the curve was found to be r = 0.9984 [$y = 0.36(\pm 0.27) + 0.79 (\pm 0.02)x$], with y being the concentration of AFB₁ (ng g⁻¹) of capers recovered and x the concentration of AFB₁ (ng g⁻¹) of capers spiked.

Moreover, a Fisher test was applied to confirm the acceptability of the linear regression. The F(1,15) ratio found

(1258.38) was greater than the critical Fisher value of 16.58 at a risk 0.1% for 1 and 15 degrees of freedom. Subsequently, the regression model can be considered acceptable. Finally, the lack of fit of the model as the experimental F ratio (1.97) is lower than the critical value F(3,15) = 9.335 with a risk a = 0.1% for 3 and 15 degrees of freedom, thus the field of linearity chosen can be approved. The mean recovery factor taken from the above equation was found to be 79% (RSD% = 6.53). The AFB₁ detection limit (DL) was determined at 0.08 ng g⁻¹ of capers and the quantification limit (QL) at 0.21 ng g⁻¹ of capers. Both DL and QL have been defined by using the signal-to-noise ratio 3:1 and 6:1, as 0.06 ng g⁻¹ and 0.12 ng g⁻¹, respectively. In the present study the lower DL = 0.06 ng g⁻¹ was used throughout this work.

3.2. Aflatoxin B₁ Production in Capers

In the present study, *A. parasiticus* was used since AFB₁ production is a more constant trait in this fungus than in *A. flavus* [38]. Moreover, AFB₁ was studied throughout this research as it is the most potent mycotoxin and usually the one produced at the highest levels by toxigenic strains [39].

To our knowledge, there is no information in the literature concerning the AFB₁ occurrence and production in capers. Hence, caper was selected as the natural product to be used as a substrate in order to study the effect of different MeJA concentrations on AFB₁ production. Moreover, capers were damaged in order to facilitate fungus penetration and achieve results in a short time.

Previously, [40] showed that MeJA at concentrations of 10⁻⁴ M and 10⁻⁶ M had no significant effect on the mycelial growth in yeast extract sucrose medium (YES) but did affect AFB₁ production. On the contrary, treatment of *A. parasiticus* cultures with 10⁻² M MeJA inhibited mycelial growth and AFB₁ production as well. It should be noted that a preliminary study of different MeJA concentrations on the *A. parasiticus* growth in AFPA medium was applied (Markaki unpublished data Fig.1), which revealed that MeJA concentrations below 10⁻³ M (corresponding to 2.267 mg MeJA) did not affect the mycelial growth of *A. parasiticus*.

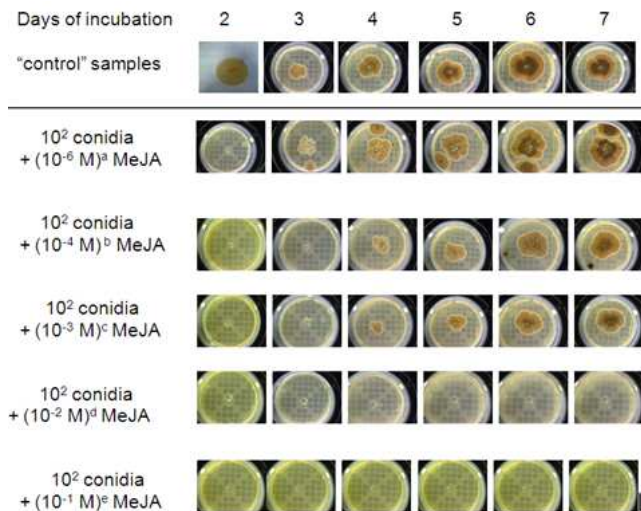


Figure 1. Macroscopic observation of the inhibition effect of MeJA on *A. parasiticus* in AFPA. The MeJA concentrations used, correspond to the MeJA concentrations added to the caper samples (ranging from 10^{-6} M to 10^{-1} M); (a: 0.0022 mg, b: 0.227 mg, c: 2.267 mg, d: 22.667 mg, e: 226.667 mg). Concentrations above 10^{-3} M MeJA significantly inhibited mycelial growth. Mycelial growth in control samples (without MeJA addition) was not affected.

Therefore, we were interested in adding MeJA at higher concentrations, such as 10^{-3} , 10^{-2} and 10^{-1} M that could inhibit AFB₁ production. The MeJA concentrations of 10^{-6} M and 10^{-4} M were additionally used in order to have comparable results of the AFB₁ production in capers against the AFB₁ production in YES medium as reported from our laboratory by [40].

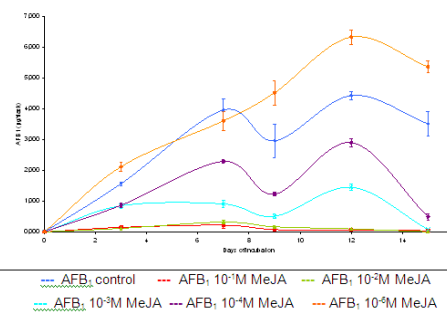


Figure 2. Aflatoxin B₁ production in capers by *A. parasiticus*. Caper samples with MeJA concentrations 10^{-1} M, 10^{-2} M, 10^{-3} M, significantly reduced AFB₁ production against control, while concentrations above 10^{-4} M stimulated AFB₁ production by *A. parasiticus* in capers.

Table 3. AFB₁ production by *A. parasiticus* in caper after MeJA addition at different concentrations

Cultures						
	caper + 10^2 conidia (control) ^a	caper + 10^2 conidia + MeJA (10^{-1} M) ^b	caper + 10^2 conidia + MeJA (10^{-2} M) ^c	caper + 10^2 conidia + MeJA (10^{-3} M) ^d	caper + 10^2 conidia + MeJA (10^{-4} M) ^e	caper + 10^2 conidia + MeJA (10^{-6} M) ^f
AFB ₁ production						
Days	µg flask ⁻¹ h (±SD)	µg flask ⁻¹ h (±SD)	µg flask ⁻¹ h (±SD)	µg flask ⁻¹ h (±SD)	µg flask ⁻¹ h (±SD)	µg flask ⁻¹ h (±SD)
0	N.D. ^g	N.D. ^g	N.D. ^g	N.D. ^g	N.D. ^g	N.D. ^g
3	1.57 (±0.06)	0.15 (±0.03)	0.12 (±0.024)	0.85 (±0.10)	0.87 (±0.08)	2.12 (±0.15)
7	3.96 (±0.35)	0.22 (±0.11)	0.32 (±0.076)	0.91 (±0.11)	2.29 (±0.06)	3.61 (±0.31)
9	2.96 (±0.55)	0.077 (±0.005)	0.17 (±0.036)	0.51 (±0.08)	1.23 (±0.07)	4.52 (±0.39)
12	4.43 (±0.13)	0.070 (±0.016)	0.10 (±0.021)	1.45 (±0.10)	2.90 (±0.13)	6.33 (±0.23)
15	3.51 (±0.40)	0.042 (±0.010)	0.008 (±0.002)	0.08 (±0.02)	0.49 (±0.10)	5.36 (±0.20)

^a caper samples inoculated with *A. parasiticus*

^b 226.667 mg MeJA 15g caper⁻¹

^c 22.667 mg MeJA 15g caper⁻¹

^d 2.267 mg MeJA 15g caper⁻¹

^e 0.227 mg MeJA 15g caper⁻¹

^f 0.0022 mg MeJA 15g caper⁻¹

^g Non-Detectable

^h Corresponds to 15g of capers flask⁻¹

The MeJA concentrations used in this work were chosen to be both above as well as below the concentration of 10^{-4} M. The latter was found to be effective as a post-harvest treatment for suppressing the decay caused by *Botrytis cinerea* on strawberries, for reducing the decay by *Penicillium digitarum* in grape fruit as well as for reducing microbial contamination in celery and peppers [41].

3.3. MeJA Effect on AFB₁ Production in Capers

In the present study, AFB₁ production was measurable from the day 0 of the incubation due to the inoculation of capers with 10^{-2} conidia of *A. parasiticus*.

Maximum AFB₁ production in samples treated with MeJA at concentrations of 10^{-6} M, 10^{-4} M and 10^{-3} M amounted to 6.33, 2.90 and 1.45 µg flask⁻¹ (15g) on the 12th day of observation, respectively. Furthermore, maximum AFB₁ output in samples treated with MeJA at higher concentrations, namely 10^{-2} M and 10^{-1} M, was 0.32 and 0.22 µg flask⁻¹ on the 7th day of observation, respectively. AFB₁ maximum production in control samples (without MeJA addition) inoculated with *A. parasiticus*, was 4.43 µg flask⁻¹ on the 12th day of observation. The AFB₁ amounts produced in samples treated with 10^{-2} M and 10^{-1} M MeJA are negligible compared to the AFB₁ production in samples with MeJA added at concentrations of 10^{-6} M, 10^{-4} M, 10^{-3} M and in control samples as well, during the entire incubation period (Fig. 2 Table 3). It must be mentioned that AFB₁ in not inoculated samples of caper either with or without MeJA addition was not detectable.

The statistical analysis using one-way ANOVA, showed that the $F_{\text{exp}} = 8.110$ was higher than the $F_{\text{theor}} = 2.533$ for df 5, 30. Thus, the AFB₁ difference between the six groups is statistically significant ($p < 0.05$).

AFB₁ production after MeJA addition at a concentration of 10^{-6} M was higher compared to control, particularly after the 9th day of observation. The t-test applied on the AFB₁ production between control samples and samples treated with 10^{-6} M MeJA showed that $t_{\text{exp}} = -2.288 < t_{\text{theor}} = 2.571$ (5 df, $p = 0.05$); Following this, the observed difference is not statistically significant for the entire incubation period. After the 9th day ($t_{\text{exp}} = 16.70 > t_{\text{theor}} = 4.303$ (2 df, $p = 0.05$)) the AFB₁ output is significantly higher compared to control. On the other hand, the AFB₁ production after MeJA addition at a concentration of 10^{-4} M was lower compared to the control samples.

The t-test $t_{\text{exp}} = 3.442 > t_{\text{theor}} = 2.571$ (5 df) showed that this reduction is statistically significant. Moreover, the AFB₁ production after addition of MeJA at concentrations of 10^{-4} M, 10^{-3} M, 10^{-2} M and 10^{-1} M was significantly lower compared to control with $t_{\text{exp}} = 3.442$, $t_{\text{exp}} = 3.667$, $t_{\text{exp}} = 3.977$ and $t_{\text{exp}} = 3.962$, respectively $> t_{\text{theor}} = 2.571$ (5 df, $p = 0.05$) for the whole period of observation.

In the samples treated with MeJA at the lowest concentrations, AFB₁ production was stimulated after the 9th day of incubation. Table 4 shows that on the 12th day of observation, AFB₁ production was increased by 42.89% in samples treated with MeJA at a concentration of 10^{-6} M, while MeJA added at a concentration of 10^{-4} M inhibited AFB₁ production by 34.54%. This is in agreement with the report on the AFB₁ production in *A. flavus* cultures treated with 10^{-4} M and 10^{-3} M MeJA by [42]. On the other hand, the highest concentrations of MeJA, 10^{-2} M and 10^{-1} M, inhibited AFB₁ by 97.74% and 98.42% on the 12th day of observation, respectively. Following that, this work concludes that the higher the MeJA concentration, the higher is the inhibition effect.

Table 4. AFB₁ Stimulation and inhibition induced in capers by different MeJA concentrations compared to control samples (without MeJA addition) on the 12th day of incubation

MeJA addition mg flask ⁻¹ ^a	AFB ₁ µg flask ⁻¹ ^a	% inhibition	% stimulation
0	4.43	—	—
0.0022 (10^{-6} M)	6.33	—	42.89
0.227 (10^{-4} M)	2.90	34.54	—
2.267 (10^{-3} M)	1.45	67.27	—
22.667 (10^{-2} M)	0.10	97.74	—
226.667 (10^{-1} M)	0.07	98.42	—

^a corresponds to 15 g of capers flask⁻¹

In the case of *Aspergilli*, interactions between jasmonates, mycelial growth and aflatoxin production have been reported by several authors. These interactions are of great interest as they suggest that there is a mechanism involving plant LOX

pathways that affect aflatoxin biosynthesis. It is interesting that both inhibition and stimulation of aflatoxin production by various LOX metabolites have been reported. Furthermore, some hydroperoxy fatty acids may exert a stronger signaling influence on aflatoxin/sterigmatocystin (AF/ST) biosynthesis than on others. For example, aflatoxin biosynthesis by *A. parasiticus* was stimulated in a synthetic medium containing a mixture of 30% 13S-HPODE and ~70% 13-HPODE although 13-HPODE had an inhibitory effect when tested alone. In addition MeJA treatment at concentrations ranging from 10^{-6} M to 10^{-3} M reduced AFB₁ production by *A. flavus* grown on either Czapek Yeast Extract Agar (CYA) medium or pistachios in storage [42].

On the contrary, [43] reported that treatment with MeJA at a concentration of 10^{-4} M stimulated AFB₁ production by *A. parasiticus* grown on YES medium. [44] underlined that lipoxygenase generated signals, such as jasmonates, have both inhibitory and promoting effects on the AFB₁ production. Moreover, [40] reported that either treatment of *A. parasiticus* cultures with 10^{-2} M MeJA or with 10^{-4} M and 10^{-6} M MeJA either inhibited or stimulated AFB₁ production, respectively.

By studying the MeJA concentrations added in caper samples used in the present study, the median inhibitory concentration (IC₅₀) was calculated at 0.433 mg MeJA flask⁻¹. This is an intermediate MeJA concentration found to be between 10^{-3} M and 10^{-4} M.

Previously, [45] showed that when olives were treated with MeJA at different concentrations, AFB₁ production was concentration-dependent in both inoculated olives with *A. parasiticus* as well as in non-inoculated olives. Meanwhile, [42], based on their observations with pistachios (a more favorable substrate for AFB₁ biosynthesis by aflatoxigenic fungi compared to olives), exhibited reduced aflatoxin contamination when exposed to MeJA.

AFB₁ biosynthesis by *A. parasiticus* is a natural fungal reaction that occurs when the death phase has approached and the secondary metabolism begins. However, when MeJA is added, differences occur in the mycotoxin biosynthesis in both a synthetic medium (YES) [40] as well as in capers. Furthermore as the mode of action of MeJA is still unclear the so called “host-fungus interaction” theory, can be used in order to explain to a certain extent our experimental results.

According to this theory, since both plant (Ja's, MeJA) as well as fungal (psi factors) oxylipins resemble each other in terms of structure and biosynthesis, it is likely that fungi may use endogenous metabolic lipid enzymes as well as endogenous oxylipins in order to initially colonize the host and subsequently to multiply and produce their toxins. Besides, recent evidence has shown that members of different fungi have evolved, in order to use the plants defense response deriving from the plant oxylipins for their own population [46,47].

In this case, the observed AFB₁ stimulation at the lowest MeJA concentration is likely to be induced by the “manipulation” of the plant regulator - which is actually the

plant's defense response - in favor of the fungus.

In the case of the AFB₁ inhibition, it is possible that the presence of high MeJA concentrations affects the physiological fungal activities by suppressing the mycelial growth in capers (Markaki unpublished data) as well as the AFB₁ biosynthesis, since according to the "reciprocal crosstalk" theory between host and fungus, plant oxylipins interact with the fungal system [48].

Furthermore, MeJA may act at a genetic level, by inducing partial or complete AFB₁ inhibition. Thus, it is essential to further study the genetic profile of the fungus with or without MeJA addition.

4. Conclusions

The results of the present study revealed the possibility of the existence of different mechanisms, by which MeJA might influence AFB₁ biosynthesis when different concentrations are used. Moreover, as it was shown in the present study, the effectiveness of MeJA produced by plants depends, in addition to other factors, on its concentration levels and therefore AFB₁ production could be predictable only under defined conditions. However, in this work, it was revealed that MeJA concentrations above 10⁻⁴ M inhibited AFB₁ production.

Thus, the use of MeJA in crop storage areas could be an effective, simple, and safe way for reducing post-harvesting aflatoxin production. This suggestion should be applied with thoughtfulness since the presence of mycotoxins in food products and the amounts produced, entirely depend on the ecological and processing parameters of the foodstuff [49].

Nomenclature

AFB₁, Aflatoxin B₁; AFB_{2a}, Aflatoxin B₁ hemiacetal derivative; A., *Aspergillus*; AFPA, *Aspergillus flavus parasiticus* agar; CFU, Colony forming units; CZA, Czapek Dox agar; HPLC, High performance liquid chromatography; SD, Standard deviation; YES, Yeast Extract Sucrose; LOX, lipoxygenase; 13S-HPODE, 13S-hydroperoxy-cis-9-trans-11-octadecadienoic acid; 13S-HPOTE, 13S-hydroperoxycis- 9, trans-11, cis-15-octadecatrienoic acid; DL, limit of detection; QL, limit of quantification; MeJA methyl jasmonate; RSD, relative standard deviation.

Conflict of Interest

We hereby confirm that the authors have no conflict of interest regarding the present work.

References

- [1] D.L. Eaton, J.D. Groopman, "The Toxicology of aflatoxins, human health, veterinary and agricultural significance" San Diego: Academic Press, 1994.
- [2] M.J. Sweeney, A.D.W. Dobson, "Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species," *Int. J. Food. Microbiol.*, 1998, vol. 43, pp.141-158.
- [3] J.W. Bennett, S.B. Christensen, "New perspectives on aflatoxin biosynthesis," *Adv. Appl. Microbiol.*, 1983, vol. 29, pp.53-92.
- [4] H.K. Abbas, "Aflatoxin and Food Safety," CRC Press: Taylor & Francis Group, 2005.
- [5] A.A. Fabbri, C. Fanelli, G. Panfili, S. Passi, P. Fasella, "Lipoperoxidation and aflatoxin biosynthesis by *Aspergillus parasiticus* and *Aspergillus flavus*," *J. Gen. Microbiol.*, 1983, vol. 129, pp.3447-3452.
- [6] J.E. Mellon, P.J. Cotty, M.K. Dowd, "Influence of lipids with and without other cottonseed reserve materials on aflatoxin B₁ production by *Aspergillus flavus*," *J. Agric. Food Chem.*, 2000, vol. 48, pp.3611-3615.
- [7] M.H. John, J. Schmidt, R. Walden, J. Shell, "Cell signaling by oligosaccharides," *Trends Plant Sci.*, 1997, vol. 2, pp.111-115.
- [8] A. Pühler, M. Arlat, A. Becker, M. Gottfert, J.P. Morrissey, F. O'Gara, "What can bacterial genome research teach us about bacteria-plant interactions?" *Curr Opin Plant Biol.*, 2004, vol. 7, pp.137-147.
- [9] E.W. Weiler, T. Albrecht, B. Groth, Z.Q. Xia, M. Luxem, H. Liss, L. Andert, P. Spengler, "Evidence for the involvement of jasmonates and other octadecanoid precursors in the tendrill coiling response of *Bryonia dioica*," *Phytochemistry*, 1993, vol. 32, pp.591-600.
- [10] H.J. Zeringue, "Effects of methyljasmonate on phytoalexin production and aflatoxin control in the developing cotton boll," *Biochem Syst Ecol.*, 2002, vol. 30, pp.497-503.
- [11] G. Agär, M. Türker, P. Batal, M.E. Erez, "Phytohormone levels in germinating seeds of *Zea mays* L. exposed to selenium and aflatoxins," *Ecotoxicology*, 2006, vol. 15, pp.443-450.
- [12] W.V. Dashek, G.C. Lewelly, "Mode of action of the hepatocarcinogens aflatoxins in plant systems: a review," *Mycopathologia*, 1983, vol. 81, pp.83-94.
- [13] J.D. Bewley, M. Black, "Seeds: Physiology of development and germination," New York: Plenum Press, 1985.
- [14] M.G. Smart, D.T. Wicklow, R.W. Caldwell, "Pathogenesis in *Aspergillus* ear rot of maize: light microscopy of fungal spread from wounds," *Phytopathology*, 1990, vol. 80, pp.1287-1294.
- [15] C. Fanelli, A.A. Fabbri, "Relationship between lipids and aflatoxin biosynthesis," *Mycopathologia*, 1989, vol. 107, pp.115-120.
- [16] C. De Luca, S. Passi, A.A. Fabbri, C. Fanelli, "Ergosterol oxidation may be considered a signal for fungal growth and aflatoxin production in *Aspergillus parasiticus*," *Food Addit Contam.*, 1995, vol. 12, pp.445-450.
- [17] J. Siedow, "Plant lipoxygenase: structure and function," *Annu Rev Plant Phys.*, 1991, vol. 42, pp.145-188.
- [18] H.W. Gardner, "Recent investigations into the lipoxygenase pathway of plants," *Biochim Biophys Acta*, 1991, vol. 1084, pp.221-239.
- [19] B.A. Vick, D.C. Zimmerman, "The biosynthesis of jasmonic acid: a physiological role for plant lipoxygenase," *Biochem Bioph Res Co.*, 1983, vol. 111, pp.470-477.

- [20] C. Inocencio, D. Rivera, F. Alcaraz, F.A. Tomas-Barberan, "Flavonoid content of commercial capers (*Capparis Spinosa*, *C. sicula* and *C. orientalis*) produced in Mediterranean countries," *Eur Food Res Technol.*, 2000, vol. 212, pp.70-74.
- [21] C. Proestos, I.S. Boziaris, G.J.E. Nychas, M. Komaitis, "Analysis of flavonoids and phenolic acids in Greek aromatic plants: Investigation of their antioxidant capacity and antimicrobial activity," *Food Chem.*, 2006, vol. 95, pp.664-671.
- [22] A. Akgül, M. Özcan, "Some compositional characteristics of capers (*Capparisspp.*) seed and oil," *Grasas Aceites*, 1999, vol. 50, pp.49-52.
- [23] B. Matthäus, M. Özcan, "Glucosinolates and fatty acid, sterol and tocopherol composition of seed oils from *Capparis Spinosa* Var. *Spinosa* and *Capparis ovata* Desf. Var. *canescens* (Coss.) Heywood," *J Agric Food Chem.*, 2005, vol. 53, pp.7136-7141.
- [24] G. Leondaritis, D. Galanopoulou, "Characterization of inositol phospholipids and identification of a mastoparan-induced polyphosphoinositide response in *Tetrahymena pyriformis*," *Lipids*, 2000, vol. 35, pp.525-532.
- [25] J.J. Pitt, "Methods for the mycological examination of food," NATO ASI Ser., 1986, No. 122.
- [26] J.E. Smith, M. Moss, "Mycotoxins: Formation, Analysis and Significance," New York: Wiley, 1985.
- [27] C.N. Shih, E.H. Marth, "Experimental production of aflatoxin on brick cheese," *J Milk Food Technol.*, 1972, vol. 35, pp.585-587.
- [28] W.O. Ellis, J.P. Smith, B.K. Simpson, H. Ramaswamy, G. Doyon, "Growth and aflatoxin production by *Aspergillus flavus* in peanuts stored under modified atmosphere packaging (MAP) conditions," *Int J Food Microbiol.*, 1994, vol. 22, pp.173-187.
- [29] R. Eltem, "Growth and aflatoxin B₁ production on olives and olive paste by molds isolated from Turkish-style natural black olives in brine," *Int J Food Microbiol.*, 1996, vol. 32, pp.217-223.
- [30] I.M. El-Refai, O.A. Awadalle, A.M. Abou Zeid, "Effect of some insect growth regulators on growth of *Aspergillus flavus* and its productivity of aflatoxin B₁ and lipids," *Food Addit Contam.*, 1995, vol. 12, pp.585-590.
- [31] D. Abramson, R.M. Clear, "A convenient method for assessing Mycotoxin production in cultures of *Aspergilli* and *Penicillia*," *J Food Prot.*, 1996, vol. 59, pp.642-644.
- [32] K.K. Sinha, A.K. Sinha, G. Prasad, "The effect of clove and cinnamon oils on growth of and aflatoxin production by *Aspergillus flavus*," *Lett Appl Microbiol.*, 1993, vol. 16, pp.114-117.
- [33] D. Leontopoulos, A. Siafaka, P. Markaki, "Black olives as substrate for *Aspergillus parasiticus* and aflatoxin B₁ production," *Food Microbiol.*, 2003, vol. 20, pp.119-126.
- [34] E. Daradimos, P. Markaki, M. Koupparis, "Evaluation and validation of two fluorometric HPLC methods for the determination of aflatoxin B₁ in olive oil," *Food Addit Contam.*, 2000, vol. 17, pp.65-73.
- [35] R.D. Stubblefield, "Optimum conditions for formation of aflatoxin M₁-trifluoroacetic acid derivative," *J Assoc Off Anal Chem.*, 1987, vol. 70, pp.1047-1049.
- [36] D.S.P. Patterson, "Structure, Metabolism and Toxicity of Aflatoxins: A Review," *Cah Nutr Diet.*, 1976, vol. 11, pp.71-76.
- [37] J. Fowler, L. Cohen, "Practical statistics for field biology," Chichester, England: John Wiley & Sons, 1997.
- [38] J. Yu, P.K. Chang, J. Cary, M. Wright, D. Bhatnagar, T.E. Cleveland, G.A. Payne, J.E. Linz, "Comparative mapping of aflatoxin pathway gene clusters in *Aspergillus parasiticus* and *Aspergillus flavus*," *Appl Environ Microbiol.*, 1995, vol. 61, pp.2365-2371.
- [39] G.B. Burow, T.C. Nesbitt, J. Dunlap, N.P. Keller, "Seed lipoxygenase products modulate *Aspergillus* mycotoxins biosynthesis," *Mol Plant Microbe In.*, 1997, vol. 3, pp.380-387.
- [40] D.M. Meimaroglou, D. Galanopoulou, P. Markaki, "Study of the effect of methyl jasmonate concentration of aflatoxin B₁ biosynthesis by *Aspergillus parasiticus* in yeast extract sucrose medium," *Int J Microbiol.*, 2009, vol. 2009, pp.1-7.
- [41] G.A. González-Aguilar, J.C. Buta, C.Y. Wang, "Methyl jasmonate reduces chilling injury, symptoms and enhances colour development of "Kent" mangoes," *J Sci Food Agr.*, 2001, vol. 81, pp.1244-1249.
- [42] M. Goodrich-Tanrikulu, N.E. Mahoney, S.B. Rodriguez, "The plant growth regulator methyl jasmonate inhibits aflatoxin production by *Aspergillus flavus*," *Microbiology*, 1995, vol. 141, pp.2831-2837.
- [43] S. Vergopoulou, D. Galanopoulou, P. Markaki, "Methyl jasmonate stimulates aflatoxin B₁ biosynthesis by *Aspergillus parasiticus*," *J Agric Food Chem.*, 2001, vol. 7, pp.3494-3498.
- [44] R.A. Holmes, R.S. Boston, G.A. Payne, "Diverse inhibitors of aflatoxin Biosynthesis," *Appl Microbiol Biot.*, 2008, vol. 78, pp.559-572.
- [45] P. Markaki, K. Velivassaki, D. Giannitsis, D. Galanopoulou, "Methyl jasmonate effect on AFB₁ production by olives inoculated with *A.parasiticus* is concentration dependent," in *Mycotoxins and Phycotoxins, Advances in determination, toxicology and exposure management*, H. Njapau, S. Trugillo, H.P. van Egmond and D.L. Park, Eds. Wageningen Academic, The Netherlands, 2006, pp.259-264.
- [46] N.C. Avanci, D.D. Luche, G.H. Goldman, M.H. Goldman, "Jasmonates are phytohormones with multiple functions, including plant defense and Reproduction," *Genet Mol Res.*, 2010, vol. 9, pp.484-505.
- [47] S.A. Christensen, M.V. Kolomiets, "The lipid language of plant-fungi interactions," *Fungal Genet Biol.*, 2011, vol. 48, pp.4-14.
- [48] M. Brodhagen, D. Tsitsigiannis, E. Hornung, C. Goebel, I. Feussner, N. Keller, "Reciprocal oxylipin-mediated cross-talk in the *Aspergillus*-seed pathosystem," *Mol Microbiol.*, 2008, vol. 67, pp.378-391.
- [49] O. Filtenborg, J.C. Frisvad, U. Thrane, "Moulds in food spoilage," *Int Food Microbiol.*, 1996, vol. 33, pp.85-102.