

Computation of electronic proprieties of the DNA and RNA bases

Mariana Virginia Popa

Electronic and Telecommunication, Autonomous University of the Hidalgo State, Mexico

Email address:

virginia_popa@yahoo.com.mx

To cite this article:

Mariana Virginia Popa. Computation of Electronic Proprieties of the DNA and RNA Bases. *International Journal of Computational and Theoretical Chemistry*. Vol. 2, No. 4, 2014, pp. 26-40. doi: 10.11648/j.ijctc.20140204.11

Abstract: In this paper the authors report the optimizations of the DNA and RNA bases (adenine, cytosine, thymine, guanine and uracil) for to determine the electronic proprieties and are employed the LSDA/6-31++G, PBE/6-31++G, PBE/LANL2DZ and PBE/SDD levels of theory both in gas phase and in the presence of the solvent water with the actual implementation of the polarized continuum model of Tomasi (PCM). And to provide the IPV, EAV, hardness, dipole moment and electronegativity (χ). The vibrational frequencies are description to purine and pyrimidine bases.

Keywords: Purine and Pyrimidine, Electronic Proprieties, DFT for DNA and RNA

1. Introduction

The importance end preoccupation in the field of the biochemistry and theoretical chemical-physics is to know the active centre in the ADN and RNA chain. On the other hand, the interaction of the metals with the ADN and RNA bases¹⁻⁹ has been the subject of numerous publications. The ab initio calculations as the properties such the electron affinities (EAs) and ionization potentials (IPs) have been presented en many works with different levels of theory and are reported experimental data also¹⁰⁻²⁵. The difference between vertical IPs, vertical EAs and their adiabatic counterparts consist by the nuclear reorganization energy on ion formation. The positive values for the electron affinities show the exothermic process and thermodynamically stable state¹⁶⁻²². It is now well established that the pyrimidine are better electron acceptors than the purine. In another publication a comprehensive and reliable computational study of the first ionization energies of the DNA and RNA bases have been calculated in the gas fase and in aqueous solution at HF and MP2 levels of theory and 6-31++G(d,p) base; upon where HF level in gas phase underestimated the ionization energies by ca. 1eV and for guanine the theoretical data is in good agreement with the experimental value in aqueous solution¹⁷. All of the calculations in order to determined vertical IPs presented by David²³ for the canonical DNA bases are based on optimization with Moller Plessetperturbational theory (MP2), or the B3LYP functional, in conjunction with a different basis sets with Polarized Continuum Model in water

and gas phase. The results are very close to the experimental data with exception when MP2 theory are used, the dates of vertical ionization potentials are overestimate. Norinder²⁴ are investigate the geometries for different tautomers of the nucleic bases using the AM1 method bat in the present work is not of interest because all structure are en stable form. Comparison of calculated and experimental value the bond length and ionization potentials are overestimated in these systems. As the DNA bases is very popular benchmark molecules the papers^[25-27] mentioned the bond length and angles, vibrational frequencies. In²⁸ the plane wave basis set in conjunction with ultrasoft pseudopotentials yield bond lengths and angles are close to conventional quantum chemistry results. In this regard all geometries of DNA bases were performed using PW91/6-311G(2df,2pd) and MP2 theory (cc-pVTZ, aug-cc-pVTZ and cc-pVQZ larger basis) used PQS Parallel Quantum Solutions, the bond length deviations are possible for the plane-wave/ultrasoft pseudopotentials method. When predict the bond length and angle for large systems it is useful to introduce PW91. For the best prediction of the reactivity to the AND and RNA bases is very important view the shape of the SOMO follows the spin density distribution of the anion radical in their neutral geometries²⁹. In the literature has written many papers where mention the influence of the charge, the HOMO and LUMO energies, the hardness, Fukui Functions condensate, and electronegativity for the better understand of the reactivity of the molecule in gas phase and different solvents. Since there are difficulties in determining the experimental

ionization potentials (IPs), adiabatic electron affinities of ADN and RNA bases and the active centers to, the goal of the present work is to perform the results employed ionization potentials and electron affinities vertical, and electronegativity, for describe the possible mechanism of reaction the adenine, cytosine, guanine and uracile with different chemical reagent. So regard than Mulliken charge are not a very good describer of the activity of the atoms in the molecule for this reason in this paper we employed the NPA and CHELPG charge to.

2. Methods

All of the calculations presented here are optimized in gas phase and aqueous medium with $\epsilon = 78.39$ (are employed Thomasi PCM model³⁰). In the present work are employed DFT method with PBE and LSDA functional in combination with several 6-31++G, SDD, LANL2DZ basis set³¹. The full optimizations of the DNA/RNA single bases guanine (G), adenine (A), cytosine (C), and uracil (U) structures are also performed with Gaussian 03W program package³². The optimizations were followed by frequency calculations at the same level and for the HOMO and spin density are visualized

with Gauss View computational program. By definition, the vertical EA (VEA) is the energy released when an electron is added to a neutral molecule and calculated as an energy difference between the anion and the neutral molecule evaluated at the geometry for the neutral parent species. The energy of vertical ionization potentials (VIP) are calculated as an energy difference between the cationic and the neutral molecule evaluated at the geometry for the neutral DNA and RNA bases.

The absolute hardness is the resistance of the chemical potential to change in the number of electrons³³.

The electronegativity (χ) of an atom or molecule is the negative of the chemical potential μ of its electronic cloud:

$$\chi = -\mu = (\partial E / \partial N)_V \quad (1)$$

Operational definitions of χ and chemical hardness (η) are provided by the finite difference approximation $\eta = (I-A)/2$, $\chi = 1/2(I+A)$, where I and A are the ionization potential and electron affinity of the species in question³⁴.

3. Results and Discussion

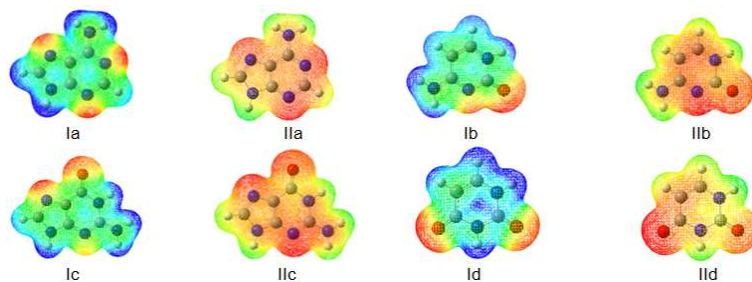


Fig 1. The surfaces of the electron density mapped with ESP (isovalue = 0.006) of the DNA and RNA bases optimized with all level of theory employed in this work in gas phase and solvent water also ($\epsilon = 78.39$) for neutral (I) and anion structure (II): a) adenine, b) cytosine, c) guanine and d) uracile.

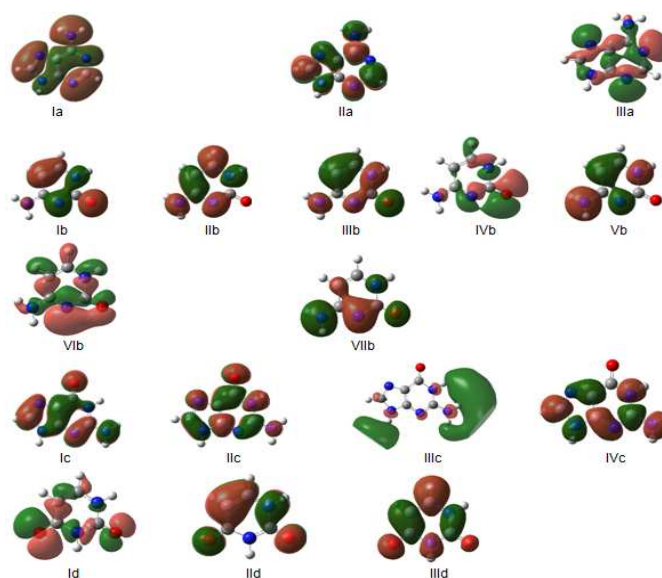


Fig 2. The HOMO and LUMO (isovalue = 0.02) of the DNA and RNA bases optimized with all level of theory employed in this work in gas phase and solvent water also ($\epsilon = 78.39$) for neutral HOMO and LUMO, anion HOMO and cation HOMObases; a) adenine, b) cytosine c) guanine and d) uracile.

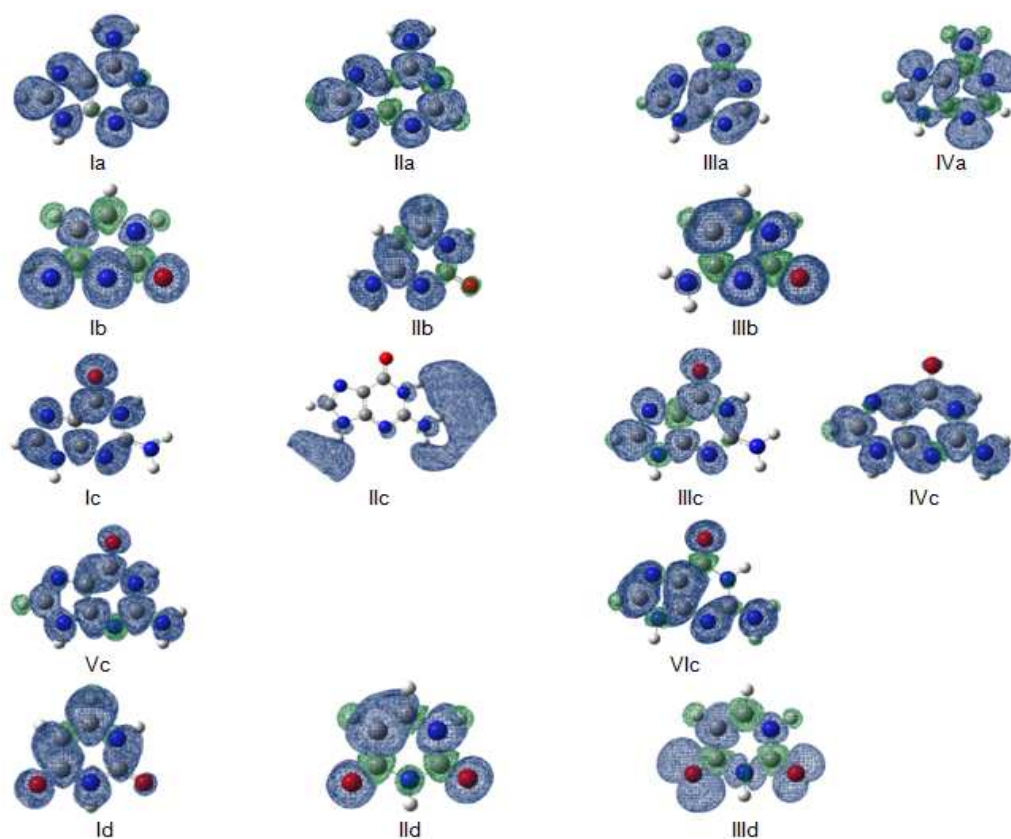
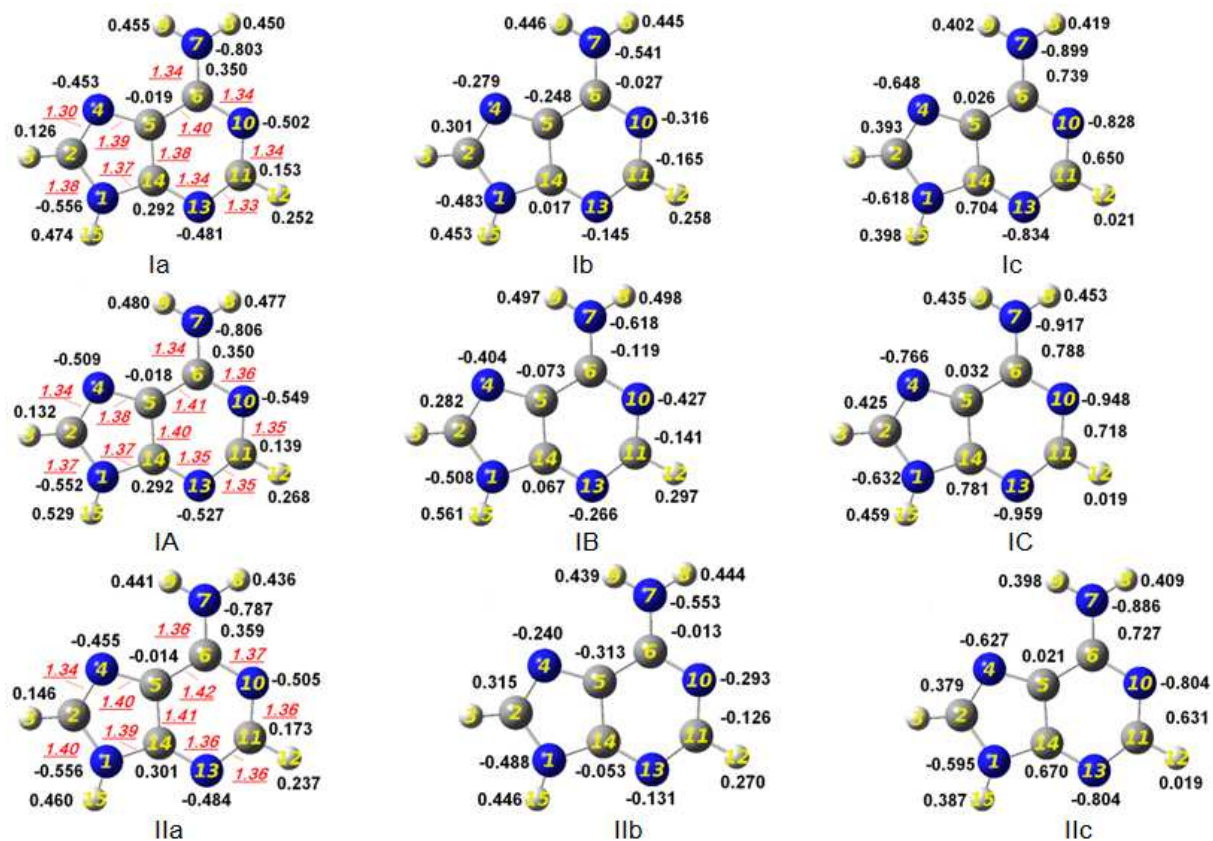


Fig 3. Spin density of anion and cation on the geometry of the optimized neutral structure (isovalue = 0.004) (IV): a) adenine, b) cytosine, c) guanine, and d) uracile.



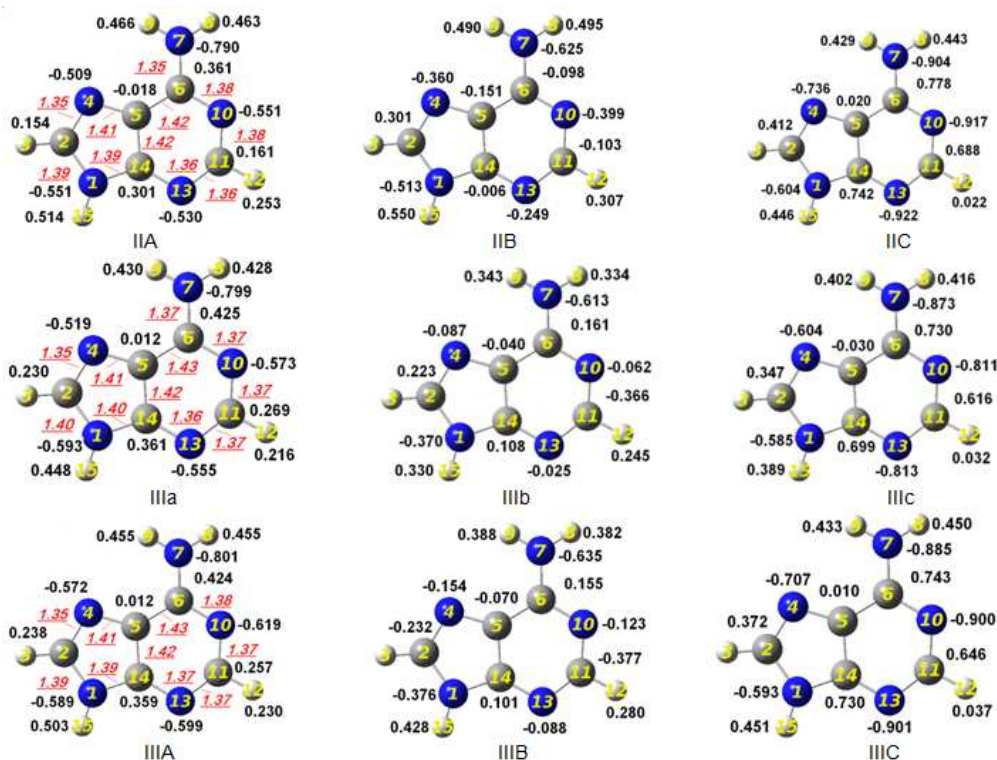
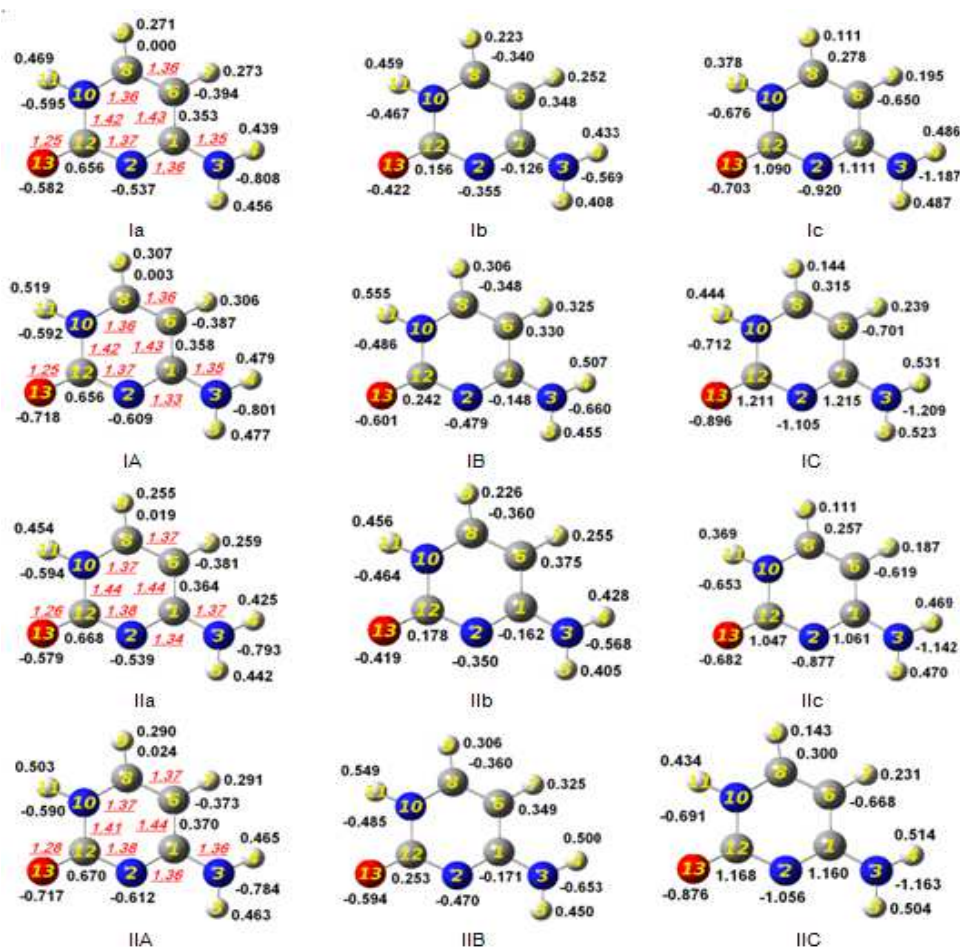


Fig 4. The charges of adenine. Bond lengths in Å - optimized with LSDA/6-31++G level of theory, II - optimized with PBE/6-31++G level of theory, III - optimized with PBE/LANL2DZ or PBE/SDD level of theory ; a -NPA, b -Mulliken, c -CHELPG charge in gas phase; A - NPA, B - Mulliken and C - CHELPG charge in solvent water ($\epsilon = 78.39$).



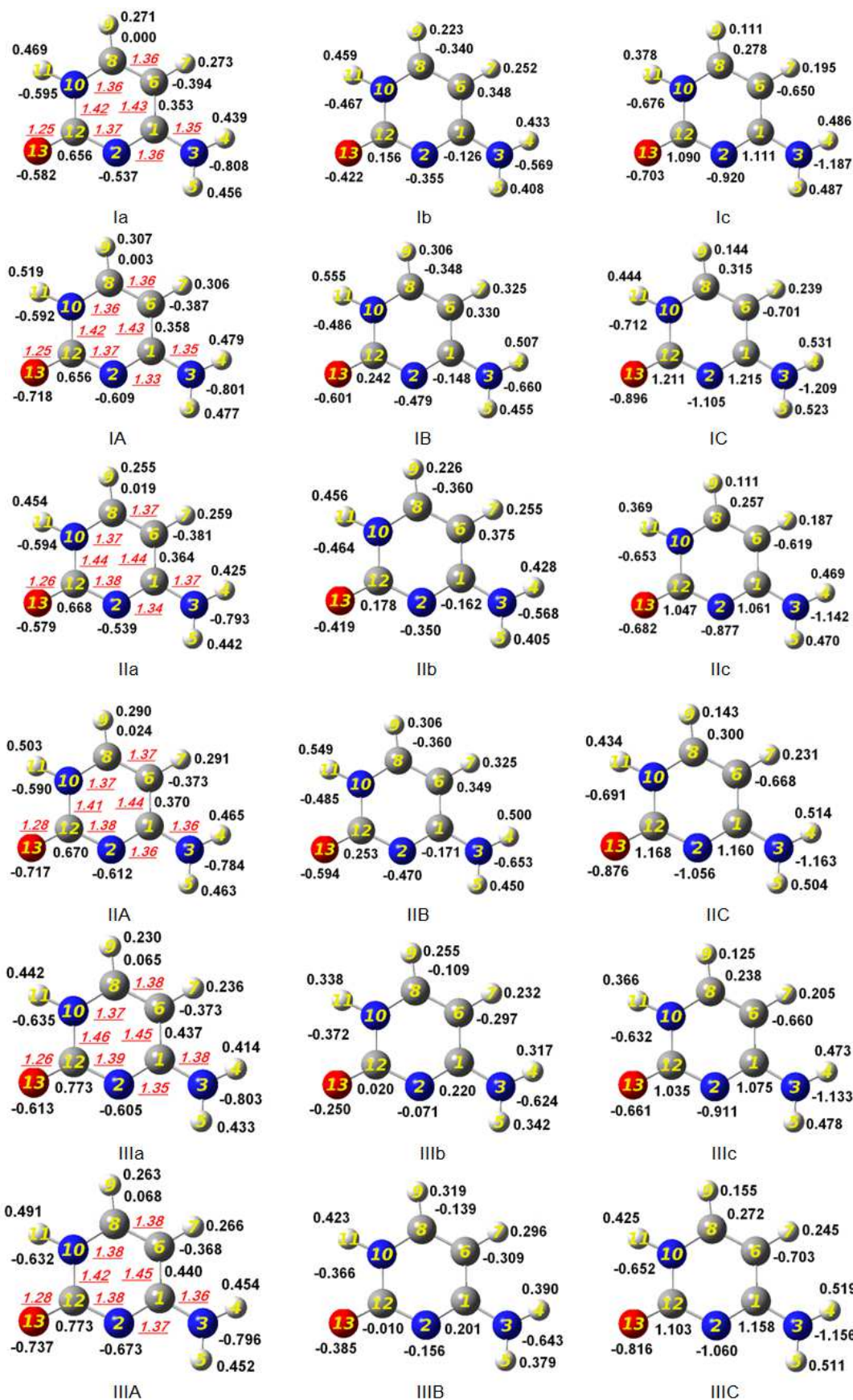


Fig 5. The charges of cytosine. Bond lengths in Å. I - optimized with LSDA/6-31++G level of theory, II - optimized with PBE/6-31++G level of theory, III - optimized with PBE/LANL2DZ or PBE/SDD level of theory; a - NPA, b - Mulliken, c - CHELPG charge in gas phase; A - NPA, B - Mulliken and C - CHELPG charge in solvent water ($\epsilon = 78.39$).

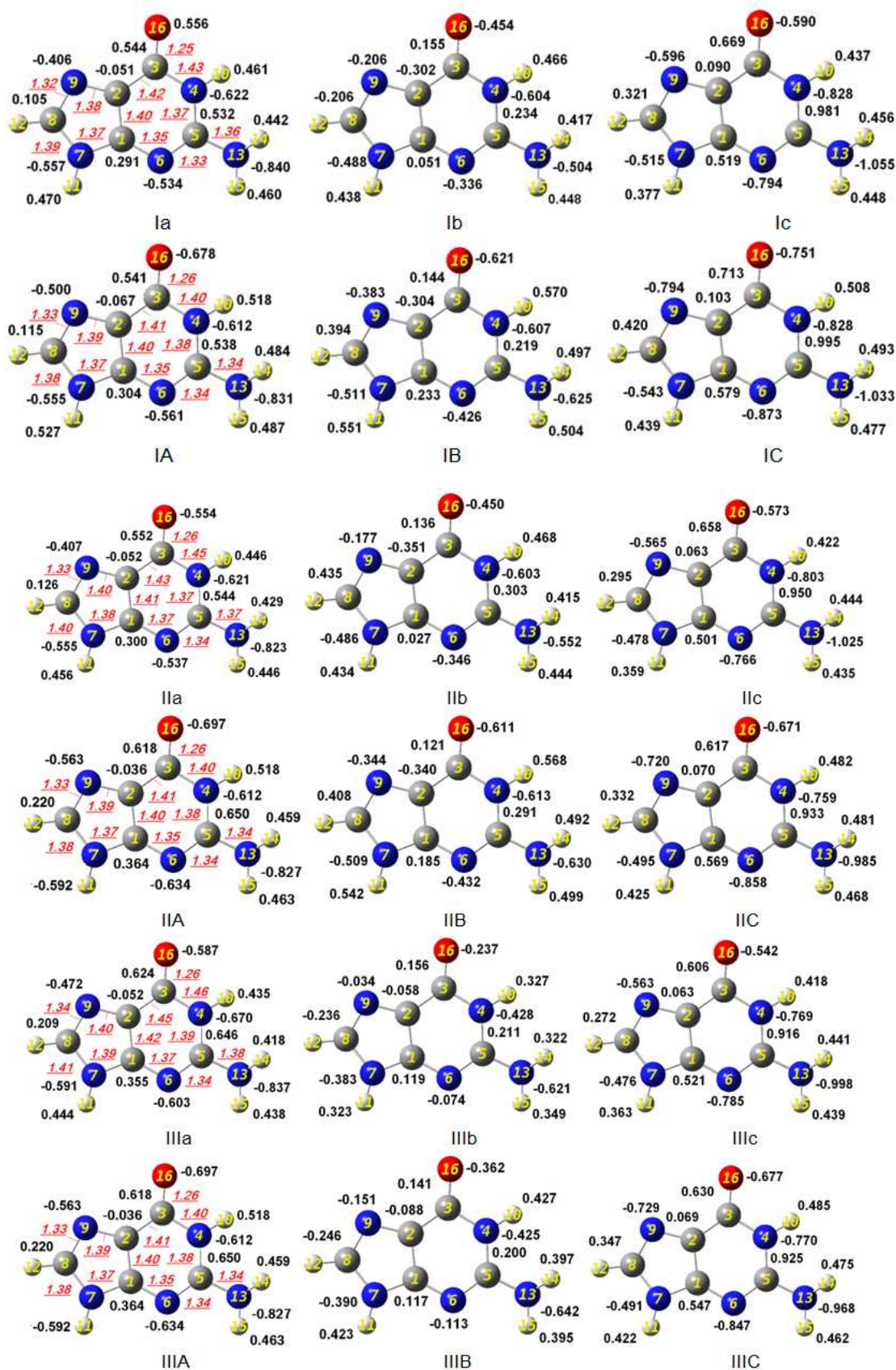
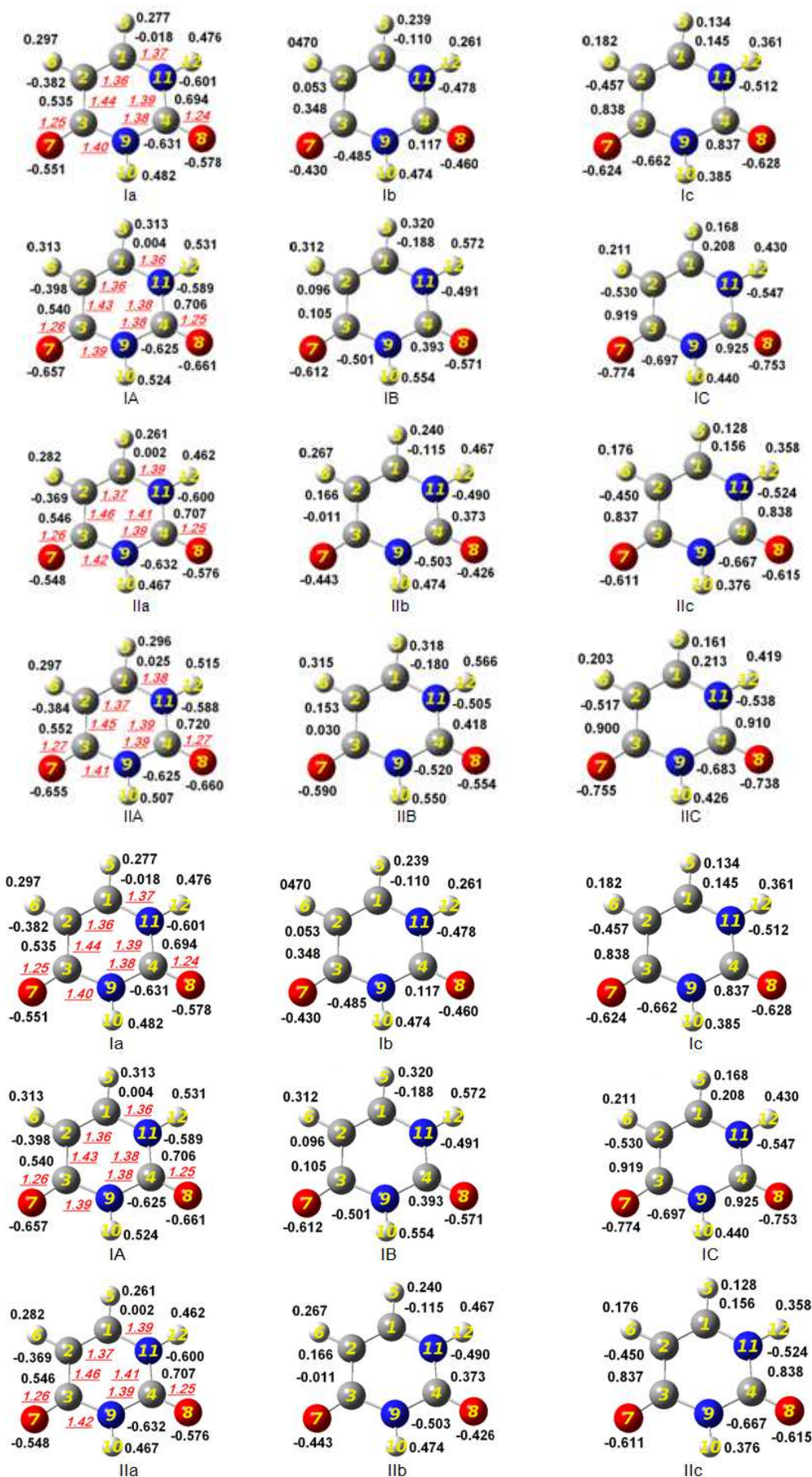


Fig 6. The charge of guanine. Bond lengths in Å. I - optimized with LSDA/6-31++G level of theory, II - optimized with PBE/6-31++G level of theory, III - optimized with PBE/LANL2DZ or PBE/SDD level of theory; a - NPA, b - Mulliken, c - CHELPG charge in gas phase; A - NPA, B - Mulliken and C - CHELPG charge in solvent water ($\epsilon = 78.39$).



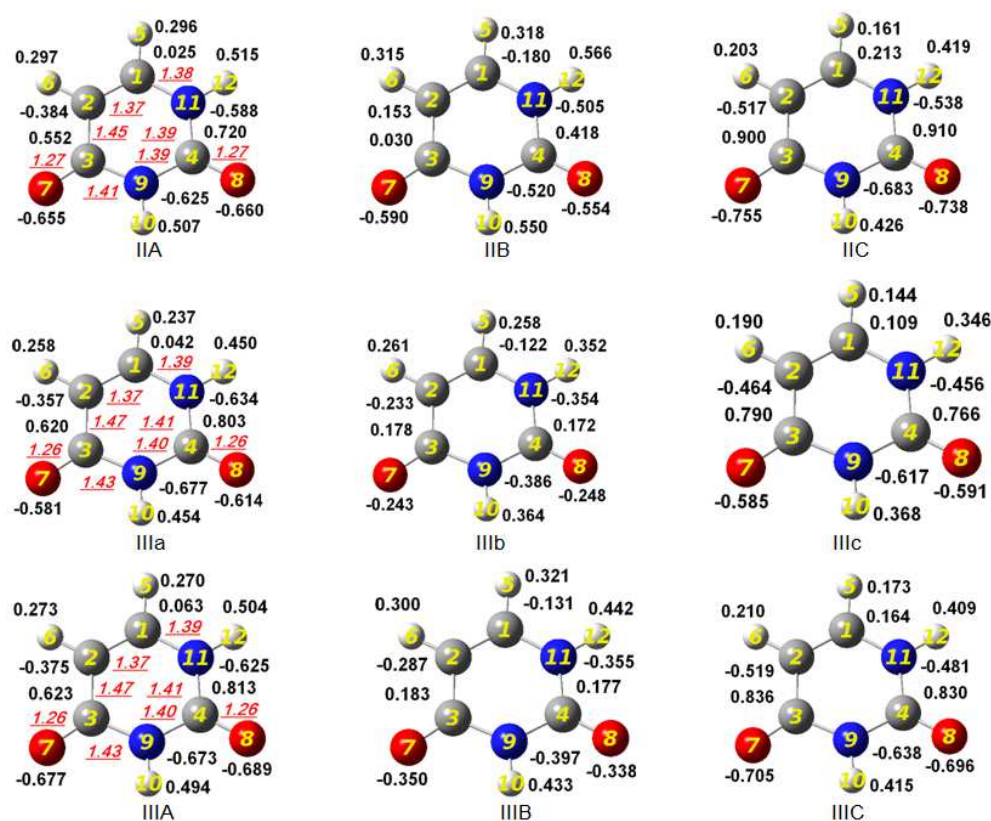


Fig 7. The charge of uracile. Bond lengths in Å. I - optimized with LSDA/6-31++G level of theory, II - optimized with PBE/6-31++G level of theory, III – optimized with PBE/LANL2DZ or PBE/SDD level of theory; a - NPA, b - Mulliken, c - CHELPG charge in gas phase; A - NPA, B - Mulliken and C - CHELPG charge in solvent water ($\epsilon = 78.39$).

The aim of this work was therefore to reinvestigate the relative stabilities of the cytosine, thymine, adenine, guanine and uracile using DFT method with full geometry optimization. This procedure has proved to be useful in the prediction of such molecular properties as the geometries, dipole moments, hardness, electronegativity, electron affinity, harmonic frequencies, the surfaces of the electron density mapped with ESP and ionization potentials, the HOMO and LUMO, spin density of the anion in the neutral structure. Although chemical reactivity is characterized by global reactivity parameters like electronegativity or hardness, the selectivity is usually understood in terms of local functions like the Fukui function $f(r)$ and local softness $s(r)$. These global and local descriptors of reactivity have been popularized within the framework of conceptual density functional theory, a field to which reviews have been dedicated recently.

The charge transfer in DNA and RNA is the fundamental interest of the reactivity by attaching or intercalating donor and acceptor chromophores electronically coupled via the bridge orbital. The interaction between electrophiles (agents with deficit of electrons) and the AND and RNA bases is a complex phenomenon. In the previous experimental investigations^{35,36} the products of the interaction of the three methylating agents characterized by increasing carcinogenic activity with ADN and ARN bases have also been examined by means of Raman and IR spectra. In this regard in the Fig. 1 I show the reactivity of adenine, cytosine, guanine, thymine

and uracile for neutral and anion structure where the red part represent the electrophilic center, the green part represent the centre of possible radical attack and the blue drawing the atoms susceptibles to nucleophilic attack. The charge is disposed above all atoms in the molecules when an electron is accepted because is possible the aromaticity to disappear, but the electrophilic centers are the same excepting for the guanine when N6 is new for the π length, Fig. 1 (Ic and Iic), Fig. 2 (IIc and IVc). N7, N10 and N4 are the most probable electrophilic center thus as has mentioned previously³⁷ employed PBE/6-31++G and LSDA/6-31++G levels of theories, and in this work Fig. 1 (Ia) for adenine. Is very know which the surfaces of the electron density mapped with ESP are not very good describer of the active centers but for larger molecules are a good method because employed Fukui Functions for the optimized structure of DNA and RNA with DFT is expensive in time.

The Fig. 2 is very useful for predict the π -conjugated molecular and delocalized frontier orbitals in the negatively molecule charged, for this reason the HOMO and LUMO can be used as indicators able to predict electrical characteristics in a first approximation of single molecules. For adenine en Fig. 2 (Ia and IIa) the HOMO and LUMO en gas phase and solvent water not change with the inclusion of the relativistic proprieties and the difference between PBE and LSDA. Remove one electron of the adenine in the HOMO denote the cation change. The cation structure of cytosine in the Fig. 2 (Vb, VIb y VIIb) are very different of the neutral and anion

structure. On the other hand, for guanine in the HOMO cation and neutral configuration Fig. 2 (Ic) are in discrepancy with the another (Figure 2 IIc) where the antibonding exist. The diffuse bases set is the possible accountable for the great contrariety regarded in the Fig. 2 (IIc and IVc). The solvent water (l) stabilize the uracile Fig. 2 (IIId, IIId) HOMO and LUMO with the HOMO cation PBE/6-31++G and PBE/SDD levels of theory exception Fig. 2 (Id). The spin surfaces of the cation Fig. 3 (IIa) with PBE and all neutral Fig. 2 (Ia) is very similar, however with LSDA/6-31++G cation, PBE/LANL2DZ cation and PBE/SDD cation in gas phase Fig. 3(IVa) and HOMO LSDA/6-31++G cation in gas phase and all cation in gas phase and solvent water Fig. 2 (IIa) are identical. For adenine the results obtained in gas phase and solvent was not found to be very important in the description of the chemical proprieties. In the literature²⁹ has been reported the SOMO surface and spin density of the anion radical of the purine and pyrimidine bases in the gas phase and including the effect of solvating (PCM model). In cytosine, Fig. 2 (Ib, IIb) with Figure 3 (IIb) are antibonding orbital but very different to SOMO surfaces and spin density of the cytosine optimized with B3LYP/D95V+D and B3LYP/6-311++G(2dp) basis sets²⁹. The HOMO LSDA/6-31++G in gas (g) phase Fig. 2 (Ib) when least expected is not different of the Fig. 2 (IIb) for the local spin density approximation in the water and PBE/6-31++G(g,l), PBE/LANL2DZ(l) and PBE/SDD(l). In Fig. 2 (Vb, VIb) HOMO PBE/6-31++G cation, HOMO PBE/LANL2DZ cation, HOMO PBE/SDD cation in gas phase and solvent water are identical but HOMO LSDA/6-31++G cation is different, Fig. 2 (VI, VII) for the local spin density (LSD) approximation.

In an unrestricted wave function, it is convenient to define a spin density $\rho^S(r)$ by³⁰

$$\rho^S(r) = \rho^\alpha(r) - \rho^\beta(r) \quad (2)$$

where electrons of α and β spin have different spatial distributions ($\rho^\alpha \neq \rho^\beta$).

It is clear that in regions of space where there is a higher probability of finding an electron of α spin then there is of finding an electron of β spin the spin density is positive. Alternatively, the spin density is negative in regions of space where electrons of β spin are most prevalent.

For adenine, Fig. 3 (Ia), the spin density is positive for the electron of α spin, when LSDA/6-31++G(g) and PBE/6-31++G(g) are used. Employed LSDA/6-31++G(l) for cation and anion, PBE/6-31++G (l) anion, PBE/LANL2DZ (g,l) for anion and PBE/SDD(g,l) for anion also in Figure 3 (IIa) the electron of β spin are un little larger the spin density that adenine structure in the Fig. 3 (Ia); with of the inclusion of diffuse functions in basis sets performed of B3LYP/D95+(D) level²⁹ adenine are the same reactivity centers.

In Fig. 3 (Ib) where are employed LSDA/6-31++G cation in water solvent N2 y N3 are when least expected the β spin greater that another atoms. The structure of the anion in Fig. 3 (IIb) is very different of the cation structure.

Fig. 2 (Ic) show the structure of the HOMO LSDA/6-

31++G(g,l), HOMO LSDA/6-31++G cation (g,l), HOMO PBE/6-31++G(g,l), HOMO PBE/6-31++G cation(g,l), HOMO PBE/LANL2DZ(g,l), HOMO PBE/LANL2DZ cation(g,l), HOMO PBE/SDD(g,l), HOMO PBE/SDD cation(g,l) when is very conspicuous the aromaticity and the π length N9-C8, N6-C5. With an electron more in anion structure to regard the anti- π length. LUMO LSDA/6-31++G(l), HOMO LSDA/6-31++G anion(l), LUMO PBE/6-31++G(l), HOMO PBE/6-31++G anion(l), LUMO PBE/6-31++G anion(l), LUMO PBE/LANL2DZ(g,l), LUMO PBE/SDD(l), HOMO PBE/SDD anion(l), HOMO PBE/LANL2DZ anion(l) in the Fig. 2(IIc) are 3 π length with 3 p orbitals alternatively disposed. The spin density of anion with the optimized structure at LSDA/6-31G(g) level of theory are similar with Fig. 2 (IIc). In HOMO PBE/LANL2DZ anion(g) Figure 2 (IVc) the π length are different disposed in comparison with the Figure2 (IIc). In the Figure 3 (IIc) where guanine is optimized with LSDA/6-31++G(g) are similar arrangement of the electron with HOMO anion in Figure 2 (IIc), it is clear that in this regions of space is a higher probability of finding an electron of α spin and for this reason the spin density is positive. In the regions of the C2 and N7 are electron of β spin when PBE/6-31++G (l), PBE/LANL2DZ (l), LSDA/6-31++G (l), PBE/SDD (l) anion (g) are used, Fig. 3 (IIIc). In Fig. 3 (IVc) the anion (g) when I performed with PBE/LANL2DZ level the contour of anion (g) HOMO, Figure 2 (IVc), is similar with the spin density, in the regions of space when are α spin is positive. In Fig. 3 (IVc) and (Vc) when I employed PBE/LANL2DZ and PBE/SDD levels of theory, in gas phase, for anion structure, the guanine are very similar with the HOMO in the Fig. 2 (IVc) and (IIc), the difference of the LANL2DZ and SDD are picture in the π length. The LANL2DZ relativist basis in Fig. 3 (IVb) are not very different in the active centers with PM3, D95V(D), 6-31G(D) and 6-31+G(D)²⁹. The 6-311++G(2dp) and 6-31++G(D) reported in literature⁹ and Fig. 3 (IIc) with diffuse functions suggest the shapes around of the N4, C5, N13, N6 and N7 atoms where the reactivity are better.

In Fig. 3 (VIc) for cation with PBE(g,l) and all bases are not difference with spin density is positive, but with LSDA/6-31++G(g,l) the distribution of the electron is the another form in the Fig. 2 (Ia).

For uracile in Figure 3 (Id) the spin density anion for all levels of theory are the exception with another purine and pyrimidine and is very different with the shape obtained employed D95+V(D). Figure 2 (Id) uracile are optimized with LSDA/6-31++G(g) for HOMO and in the water and gas phase for cation, for PBE/6-31++G and PBE/SDD are the same situation when the orbital of the O are better reactivity; LANL2DZ in water with PBE are not the same contour. The reactivity are specifically for cation with LSDA/6-31++G(l) and PBE/6-31++G(g), in O atoms, is describe in this manner how in regions of space of spin density, Figure 3 (IIIId).

The π length C3-C2-C1 are the difference HOMO and LUMO in water for PBE/6-31++G(l), HOMO PBE/LANL2DZ(l) neutral form and cation, in Figure 2 (IIId) and Figure 2 (IIIId), HOMO PBE/SDD (l) in Figure 2 (IIId) is characteristic for neutral form.

The difference in the shape of the spin density within purine and pyrimidine (uracile) are in the fact the anion are describe in only figure and for LANL2DZ and SDD the active centre are the same.

N7 are the nucleophile site for adenine with -0.803, -0.541, -0.899 for NPA, Mulliken and CHELPG charge in gas phase; -0.806, -0.618, -0.917 in solvent water when are optimized with LSDA/6-31++G; N1 and N10, in the same figure (Ia and Ib), are the second and third sites in gas phase and N13 and N1 in solvent water, Figure 4 (Ic). N10 and N13 are the probable site obtained with Fukui Function³¹ and experimental data³²⁻³⁶.

En la literatura³¹ for the electrophilic site the active site N1, C2 and C5 are not the same with Figure 4 (Ia, Ib, Ic, IA, IIB and IIIC), C5 are the active site of radical obtained with Fukui Function³¹ and in this work, Figure 4 (Ia, Ic, IA, IB and IC) in the solvent water.

With PBE/6-31++G level of theory the active centers are the same, but in solvent water are N13, N10 and N7. The LANL2DZ and SDD relativist set bases are not influence in the results, N7, N1, N10 when the difference of the SDD are (0.001, 0.001, 0.001), for Mulliken charge N7, N1, N11 (0.003, 0.006, 0.018), CHELPG N7, N13, N10 (0.001, 0, 0.002).

For cytosine in Figure 5 (Ia, Ib, Ic, IA, IB and IC) N3, O13, N2 and N10 are the active centers for the electrophilic agents when the positions of the atoms is different with the level of theory and solvent water the exceptions being when the CHELPG charge are used. N3, N10 and O13 are the active centers when the PBE/6-31++G level of theory are employed, but when solvent water are used the active centers are N3, N2 and O13, Figure 5 (IIA, IIB and IIC). The PBE/LANL2DZ or PBE/SDD level of theory are not influencing for the charge value, Figure 5 (IIIA, IIIB and IIIC) comparing with the PBE/6-31++G level. The difference between LANL2DZ and SDD for NPA charge N3, N10, O13 are (0.001, 0.001, 0.001); for Mulliken charge N3, N10, C6 are (0.003, 0.002, -0.011); for CHELPG N3, N2, O13 are (0, -0.001, 0), (-0.001, -0.001, 0) in gas phase; for NPA charge N3, O13, N2 are (0.001, 0.001, 0.001), for Mulliken charge N3, O13, N10 (0.002, 0.003, 0.001), CHELPG charge are N3, N2, O13 (-0.001, -0.002, 0) in solvent water. O13, C6, N2 are the active centers reported in literature³¹ for LSDA/6-31++G.

When increasing the basicity in order $T \leq A < C < G$ the active centers in gas phase are N13, N4, N7 and in solvent water N13, O16 and N4 Figure 6 (Ia and IA).

In Figure 6 (Ib) and (IB) N4, N13, N7 in gas phase is

different of the value obtained in solvent water N13, O16, N4. When are used CHELPG the active centers not change in solvent water, N13, N4, N6 in gas phase and N13, N6 and N4 in solvent water. Only when the water are present the O16 to put in common the electrons but the N13 strive with he. With PBE/6-31++G the active centers are close with the previously level of theory. For guanine the solvent water when are used LANL2DZ or SDD the active centers obtained with Mulliken (exception O16) and CHELPG are not change, Figure 6 (Ia, Ib, Ic, IA, IB and IC), but for the value obtained with CHELPG are the little difference (0.017, 0.011, -0.011) for LANL2DZ and SDD. The difference between LANL2DZ and SDD for NPA charge are N13, N4, N6 (0.001, 0.001, 0.001), for Mulliken charge are N13, N4, N7 (0.004, -0.004, 0), for CHELPG charge are N13, N6, N4 (0, -0.001, -0.002) in gas phase; for NPA charge are N13, O16, N6 (0.001, 0, 0.001)

In the uracile with LSDA/6-31++G and PBE/6-31++G are not neither difference between NPA, Mulliken in gas phase but in solvent water the O8, O7, N9 are the nucleophylic centers. When I optimized with PBE/LANL2DZ are the difference for the active centers (0.001, 0, 0.001), (-0.004, -0.002, 0.003), (-0.003, -0.001, 0) in gas phase and (0.001, 0.001, 0.001), (-0.005, -0.002, 0.002), (-0.001, -0.001, -0.002) in solvent water with PBE/SDD. The differences of the bond lengths are very little for all structure.

For the calculation of the IPv in this work I take into consideration the stabilization energy of the ejected hydrated electron, the experimental ground-state of the “quasi-free” electron in the liquid (-1.3 eV^{37y}) was added in the calculation of the IPv according to the equation:

$$IP_v = E_n - E_t + V \quad (3)$$

where E_n is the total energy of the neutral molecule, E_t is the total energy of the radical cation in the condensed phase, and V is the hydrated electron stabilization energy as defined above.

The bulk solvent polarization effects on the IPv and Ev are of the DNA/RNA bases, are presented in Table 2.

It was found that isolated nucleobases can stabilize the surplus electron in a dipole-bound state. In Table 7 I listed the dipole moments of the DNA and RNA bases when cytosine and uracil which have been calculated using the LSDA/6-31++G level of theory are underestimated compared to the experimental value³⁷.

Table 1. Electronic proprieties of the DNA and RNA bases in the gas phase.

Proprieties	Ip v % (eV)				Eav % (eV)					
	6-31++G ^a	LANL2DZ ^a	SDD ^a	631++G ^b	Exp ^c	6-31++G ^a	LANL2DZ ^a	SDD ^a	631++G ^b	Exp ^d
Adenine	-2.61	9.95	11.73	16.00	8.44	88.9	51.85	51.8	-29.6	-0.54
Cytosine	-4.36	7.49	-5.37	2.80	8.94	306.3	521.88	165.6	-240.6	-0.32
Guanine	-4.98	-5.70	-5.83	1.58	8.24	-115.2	260.87	300.0	-115.2	-0.46
Uracil	10.63	9.68	-2.42	6.21	9.50	81.8	95.45	100.0	-263.6	-0.22

Table 1. Continued

Proprieties	Hardness (eV)				Electronegativity (eV)			
Method employed/basis set	6-31++G ^a	LANL2DZ ^a	SDD ^a	631++G ^b	6-31++G ^a	LANL2DZ ^a	SDD ^a	631++G ^b
Adenine	4.62	5.05	5.13	5.09	3.60	4.23	4.31	4.71
Cytosine	4.92	5.80	4.66	4.37	3.62	3.81	3.80	4.82
Guanine	3.88	4.71	4.80	4.15	3.95	3.05	2.96	4.22
Uracil	5.46	5.43	4.85	4.86	5.06	4.99	4.42	5.22

^a – all of the calculations are optimized with the PBE level^b – all of the calculations are optimized with the LSDA level^c – experimental data [14]^d – experimental data [12]Table 2. Electronic proprieties of the DNA and RNA bases in the solvent ($\epsilon = 78.39$).

Proprieties	I _{pv} (eV)					E _{av} (eV)			
Método / Set de Bases	6-31++G ^a	LANL2DZ ^a	SDD ^a	6-31++G ^b	expct	6-31++G ^a	LANL2DZ ^a	SDD ^a	6-31++G ^b
Adenine	4.84	4.79	4.78	5.42	5.0	1.43	1.30	1.29	2.11
Cytosine	5.13	5.04	5.04	6.16	5.5	1.44	1.26	1.26	2.20
Guanine	4.49	4.41	4.40	5.03	4.8	1.19	0.97	0.96	1.91
Uracil	5.50	5.41	5.40	8.40	-	1.75	1.57	1.57	2.28

Table 2. Continued

Proprieties	Hardness (eV)				Electronegativity (eV)			
Método / Set de Bases	6-31++G ^a	LANL2DZ ^a	SDD ^a	6-31++G ^b	6-31++G ^a	LANL2DZ ^a	SDD ^a	6-31++G ^b
Adenine	1.71	1.74	1.74	1.66	3.13	3.04	5.26	3.77
Cytosine	1.85	1.89	1.89	1.98	3.29	3.15	3.80	4.18
Guanine	1.65	1.72	1.72	1.56	2.84	3.30	3.76	3.47
Uracil	1.87	1.92	1.92	3.06	3.62	3.49	4.14	5.34

^a – all of the calculations are optimized with the PBE level^b – all of the calculations are optimized with the LSDA level^c – experimental dataTable 3. Harmonic frequencies (cm^{-1}) of the DNA and RNA are optimized with LSDA/6-31++G.

Nr.	adenine	expt	cytosine	expt	guanine	uracil
1	165	162	138	197	137	172
2	216	214	202	232	177	183
3	265	242	349	260	200	376
4	304	276	429		314	421
5	515	298	480	330	316	516
6	526	503	518		331	534
7	562	514	543	537	361	554
8	571	521	573	575	486	654
9	589	528	601	614	523	706
10	614	566	684	637	542	724
11	649	610	721		626	779
12	668	655	772	781	628	784
13	707	672	777	818	648	816
14	724	678	808		655	961
15	818	802	926		667	968
16	843	848	954		717	1086
17	883	869	983		734	1204
18	937	927	1067	1090	793	1266
19	940	958	1117		810	1358
20	996	1005	1221	1192	826	1384
21	1084	1037	1273	1244	939	1415
22	1146	1103	1349	1337	1016	1480
23	1234	1181	1426	1422	1062	1650

Nr.	adenine	expt	cytosine	expt	guanine	uracil
24	1251	1240	1523	1475	1079	1708
25	1328	1268	1550	1539	1140	1736
26	1358	1328	1613	1595	1166	3172
27	1378	1340	1680	1656	1290	3215
28	1398	1374	1714	1720	1330	3512
29	1436	1419	3161		1354	3556
30	1467	1448	3183		1387	
31	1450	1482	3524	3441	1435	
32	1591	1584	3533		1481	
33	1629	1606	3673	3565	1543	
34	1684	1633			1585	
35	3164	3041			1622	
36	3239	3057			1656	
37	3523	3442			1747	
38	3586	3499			3240	
39	3666	3557			3492	
40					3550	
41					3583	
42					3687	

Table 4. Harmonic frequencies (cm^{-1}) of the DNA and RNA are optimized with PBE/6-31++G.

Nr.	adenine	expt	cytosine	expt	guanine	uracil
1	163	162	136	197	132	168
2	214	214	198	232	166	177
3	270	242	337	260	191	369
4	297	276	413		276	413
5	508	298	427	330	305	511
6	519	503	497		317	525
7	530	514	521	537	355	544
8	556	521	553	575	471	637
9	560	528	576	614	494	698
10	603	566	665	637	501	709
11	621	610	711		599	756
12	651	655	730	781	614	759
13	682	672	751	818	632	804
14	702	678	768		640	945
15	801	802	885		661	953
16	824	848	939		690	975
17	867	869	942		708	1072
18	921	927	1026	1090	768	1183
19	931	958	1099		792	1237
20	983	1005	1203	1192	794	1346
21	1067	1037	1215	1244	903	1378
22	1114	1103	1331	1337	992	1391
23	1212	1181	1392	1422	1018	1458
24	1227	1240	1484	1475	1054	1611
25	1285	1268	1501	1539	1087	1661
26	1315	1328	1608	1595	1132	1684
27	1342	1340	1643	1656	1266	3174
28	1370	1374	1658	1720	1307	3217
29	1398	1419	3160		1318	3524
30	1434	1448	3191		1337	3567
31	1463	1482	3539	3441	1385	
32	1572	1584	3542		1441	
33	1599	1606	3700	3565	1489	
34	1645	1633			1548	
35	3172	3041			1583	
36	3243	3057			1629	
37	3534	3442			1690	
38	3590	3499			3238	
39	3676	3557			3506	
40					3564	
41					3592	
42					3714	

Table 5. Harmonic frequencies (cm^{-1}) of the DNA and RNA are optimized with PBE/LANL2DZ.

Nr.	adenine	expt	cytosine	expt	guanine	uracil
1	159	162	136	197	132	163
2	209	214	198	232	166	175
3	258	242	337	260	191	360
4	295	276	413		276	408
5	498	298	427	330	305	497
6	509	503	497		317	515
7	515	514	521	537	355	534
8	546	521	553	575	471	629
9	554	528	576	614	494	698
10	594	566	666	637	501	708
11	642	610	712		599	744
12	652	655	729	781	614	772
13	672	672	751	818	632	807
14	693	678	768		640	933
15	796	802	884		661	948
16	823	848	939		690	960
17	847	869	942		708	1063
18	903	927	1025	1090	768	1174
19	932	958	1098		792	1229
20	968	1005	1202	1192	794	1335
21	1064	1037	1215	1244	903	1367
22	1105	1103	1330	1337	992	1379
23	1201	1181	1393	1422	1018	1444
24	1220	1240	1483	1475	1054	1608
25	1289	1268	1501	1539	1087	1664
26	1313	1328	1607	1595	1132	1681
27	1333	1340	1642	1656	1266	3176
28	1366	1374	1658	1720	1307	3220
29	1390	1419	3160		1318	3526
30	1433	1448	3191		1337	3567
31	1456	1482	3539	3441	1385	
32	1560	1584	3542		1441	
33	1581	1606	3699	3565	1489	
34	1633	1633			1548	
35	3166	3041			1583	
36	3237	3057			1629	
37	3541	3442			1690	
38	3595	3499			3238	
39	3696	3557			3506	
40					3564	
41					3592	
42					3714	

Table 6. Harmonic frequencies (cm^{-1}) of the DNA and RNA are optimized with PBE/SDD.

Nr.	adenine	expt	cytosine	expt	guanine	uracil
1	159	162	136	197	132	163
2	209	214	198	232	166	175
3	257	242	337	260	191	360
4	295	276	413		274	407
5	498	298	426	330	305	497
6	509	503	497		316	515
7	513	514	521	537	355	533
8	546	521	553	575	471	628
9	553	528	575	614	493	697
10	594	566	664	637	501	707
11	642	610	710		598	744
12	651	655	729	781	614	772
13	671	672	750	818	632	806
14	693	678	767		640	933
15	796	802	884		660	948
16	823	848	939		690	960
17	846	869	942		708	1062
18	903	927	1026	1090	768	1174
19	932	958	1098		792	1229
20	968	1005	1203	1192	794	1335

Nr.	adenine	expt	cytosine	expt	guanine	uracil
21	1064	1037	1215	1244	903	1367
22	1105	1103	1331	1337	992	1379
23	1201	1181	1392	1422	1018	1444
24	1219	1240	1484	1475	1054	1608
25	1288	1268	1501	1539	1086	1664
26	1313	1328	1608	1595	1131	1683
27	1333	1340	1642	1656	1265	3176
28	1366	1374	1659	1720	1307	3221
29	1390	1419	3161		1318	3526
30	1433	1448	3192		1337	3567
31	1456	1482	3538	3441	1385	
32	1560	1584	3542		1441	
33	1581	1606	3699	3565	1490	
34	1633	1633			1548	
35	3167	3041			1583	
36	3237	3057			1630	
37	3541	3442			1691	
38	3595	3499			3239	
39	3696	3557			3505	
40					3564	
41					3591	
42					3714	

Table 7. Dipole moments (debye) of the DNA and RNA bases

Level of theory/bases set	Adenine neutral/anion	Cytosine neutral/anion	Guanine neutral/anion	Uracil neutral/anion
LSDA/6-31++G	2.58(3.14)	7.30(5.42)	7.56(12.74)	4.9(3.71)
PBE/6-31++G	2.52(3.46)	7.20(5.77)	7.38(4.44)	4.84(5.18)
PBE/LANL2DZ	2.59(2.14)	7.14(5.15)	7.43(4.08)	4.75(3.84)
PBE/SDD	2.59(2.14)	7.13(5.14)	7.42(6.24)	4.75(3.83)
LSDA/6-31++G ($\epsilon = 78.39$)	3.79(3.74)	11.3(8.81)	11.43(14.70)	7.13(6.42)
PBE/6-31++G ($\epsilon = 78.39$)	3.72(3.62)	11.03(8.76)	11.23(14.56)	7.07(6.45)
PBE/LANL2DZ ($\epsilon = 78.39$)	3.77(3.75)	10.55(8.12)	10.93(13.96)	6.74(5.93)
PBE/SDD ($\epsilon = 78.39$)	3.77(3.76)	10.54(8.11)	10.93(13.96)	6.73(5.93)
experimental data ^x				

^x –in gase phase

4. Conclusions

N7, N10 and N4 are the most probable electrophilic center in adenine for the binding distances. The relativistic properties do not change the reactivity of the molecule. The cation structure of cytosine are very different of the neutral and anion structure. For adenine the spin density is positive for the electron of α spin when LSDA/6-31++G(G) and PBE/6-31++G(g) are used. In the water the results are better. The spin orbitals are very different of the HOMO and LUMO because are employed de 2 electron of the last orbital.

References

- [1] Prashant Jain, Pradeep T., *Biotechn. and Bioeng.* 2005, 90, 1, pp. 59-63.
- [2] Ivan Sondi, Branka S. Sondi, *J. Colloidal Interface Science* 2004, 275, pp. 177-182.
- [3] Russo N., Toscamo M., Grand A., *J. Am. Chem. Soc.* 2001, 123, pp. 10272.
- [4] Shuxi Dai, Xingtang Zhang, Tianfeng Li, et al., *Applied Surface Science* 2005, 249, 1-4, pp. 346-353.
- [5] Famulari A., Moroni F., Sironi M., Gianinetti E., Raimondi M., *J. Molec. Struct.(Theochem)* 2000, pp. 529, 209.
- [6] Ford G.P., Scribner J.D., *Chem. Res. Toxicol.* 1990, 3, pp. 219.
- [7] Cardona J.P., Lippard S.J., Gait M.J., Singh M., *J. Am. Chem. Soc.* 1982, 104, pp. 5793.
- [8] Basch H., Krauss M., Stevens W.J., Cohen D., *Inorg. Chem.* 1986, 25, pp. 684.
- [9] Nakayama N., Tanaka S., Kikuchi O., *J. Theor. Biol.* 2002, 215, pp. 13.
- [10] J.R. Wiley, J.M. Robinson, S. Ehdai, E.C.M. Chen, E.S.D. Chen, W.E. Wentworth, *Biochem. Biophys. Res. Comm.* 1991, 180, pp. 841.
- [11] E.C.M., Chen, E.S.D., W.E. Wentworth, *Biochem. Biophys. Res. Commun.* 1990, 171, 97.
- [12] K. Aflatoon, G.A. Gallup, P.D. Burrow, *J. Phys. Chem. A.* 1998, 102, pp. 6205.
- [13] V.M. Orlov, A.N. Smirnov, Ya M. Varshavsky, *Tetrahedron Lett.* 1976, 17, pp. 4377-4378.
- [14] N.S. Hush, S.C. Agnes, *Chem. Phys. Lett.* 1975, 34, pp. 11.
- [15] D.S. Michael, B. Brent, C.A.-Odile, *J. Phys. Chem.* 1994, 99, pp. 1060.
- [16] A.-O. Colson, B. Brent, M.D. Sevilla, *J. Phys. Chem.* 1993, 97, pp. 13852.

- [17] Carlos E. Crespo-Hernández, R. Arce, Y. Ishikawa, L. Gorb, J. Leszczynski, D.M. Close, *J. Phys. Chem. A*. 2004, 108, pp. 6373-6377
- [18] A. Kumar, M.K. Mohammady, P.C. Mishra, S. Suhai, *J. Comput. Chem.* 2004, 25, pp. 1047.
- [19] X Lee, Z. Cai, M.D. Sevilla, *J. Phys. Chem A* 2002, 106, pp. 9345.
- [20] S.S. Wesolowski, M.L. Leininger, P.N. Pentchev, H.F. Schaefer, *J. Am. Chem. Soc.* 2001, 123, pp. 4023.
- [21] S.D. Wetmore, R.J. Boyd, L.A. Enksson, *Chem Phys. Lett.* 2000, 322, pp. 129.
- [22] M.D. Sevilla, B. Bosler, A.O. Colson, *J. Phys. Chem.* 1995, 99, pp. 1060.
- [23] David M. Close, *J. Phys. Chem. A*. 2004, 108, pp. 10376-10379.
- [24] Ulf Norinder, *J. Molecular Structure (Theochem)* 1987, 151, pp. 259-269.
- [25] Preuss M., Schmidt W.G., Seino K., Furthmuller J., Bechstedt F., *J. Comput. Chem.* 2004, 25, pp. 112.
- [26] Fogarasi G., *J. Phys. Chem A*, 2002, 106, pp. 1381.
- [27] Podolyan Y., Rubin Y.V., Leszczynski J., *J. Phys. Chem. A* 2000, 104, pp. 9964.
- [28] Peter Pulay, Svein S., Massimo M., Jon Baker, *J. Comput. Chem.* 2005, 26 (6), pp. 599-605.
- [29] Xifeng Li, Zhongli Cai, Michael D. Sevilla, *J. Phys. Chem. A*, 2002, 106, pp. 1596-1603.
- [30] Attila Szabo, Neil S. Ostlund, "Modern Quantum Chemistry", Dover Publications, Inc., Mineola, New York, 1996, pp. 212
- [31] M. Virginia Popa, *Rev. Mex. Fis.* 2007, 53(4), pp. 241-253.
- [32] Irma N. Kolomietset al., *J. Molec. Struct.*, 1991, 250(1), pp. 1-11.
- [33] Hye-Young H. Kim, M. Cooper, Lubomir V.N., Constance M.H. and Thomas M.H., *Chem. Res. Toxicol.* 2001, 14(9), pp. 1306-1314.
- [34] Gareth K. Forde et al., *J. Phys. Chem. A*, 2006, 110(69), pp. 2308-2313.
- [35] Marzili L.G., Kistenmacher and Eichhorn G.L., (Ed. John Wiley Sons, New York, 1980) pp. 179.
- [36] Fan J-Y, Terrel M. and Denny W.A., *Anti-Cancer Drug Design* 1997, 12(4), pp. 277-293.
- [37] J. Schiedt, R. Weinjau, D.M. Neumark, E.W. Schlang, *Chemical Physics* 1998, pp. 511-524.
- [38] Netmore S.D., Boyd R. J., Enksson L.A., *Chem. Phys. Lett.*, 322, 129, pp. 1997.