
Effect of the Nutrient Composition of Biodegraded Sweet Orange (*Citrus sinensis*) Fruit Peel on the Growth Performance of Starter Broiler Chicks

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Abstract: A twenty-eight (28) day feeding trial was conducted to determine the nutrient composition of biodegraded sweet orange (*Citrus sinensis*) fruit peel and its potential as an energy source in the nutrition of starter broiler chicks. Rumen content was collected from freshly slaughtered cattle and, fresh sweet orange fruit peels (SOP) were collected from orange fruit retailers. The rumen content was mixed with water at ratio 1kg: 1L and sieved to obtain rumen filtrate (RF). The fresh sweet orange peels were divided into four equal parts in weight; T₁, T₂, T₃ and T₄, and soaked in the rumen filtrate at ratio 1 kg: 1 L in air-tight bags for 12 h, 24 h, 36 h and 48 h, respectively. The biodegraded sweet orange peels (BSOP) were sun-dried to a moisture level of about 10%, milled and each used to replace 30% of maize in the control diet (CD) to obtain broiler starter test diets T₁D, T₂D, T₃D and T₄D, respectively. Chemical analyses were carried out to determine their proximate composition and fibre fractions while, metabolizable energy content was calculated. The results showed that BSOP contained CP, CF, EE, Ash, NFE and metabolizable energy in the range of 6.78%-7.30%, 10.36%-12.30%, 1.88%-2.65%, 7.79%-11.76%, 66.04%-72.46% and 2829.44 kcal/kg-3037.97 kcal/kg, respectively. The BSOP had ADF, NDF, ADL, hemicellulose and cellulose in the range of 19.50%-22.50%, 52.30%-56.70%, 6.80%-8.40%, 32.80%-34.20% and 12.90%-14.40%, respectively. A total of one hundred and eighty day old broiler chicks (Ross 308) were randomly assigned to five dietary treatments replicated three times with equal number and similar weights in a completely randomized design. The experimental diets had significant (p<0.05) on the final weight, weight gain feed intake, feed conversion ratio, protein intake, protein efficiency ratio and mortality. The birds fed the BSOP based diets had similar non-significant (p>0.05) and inferior values to the birds on the maize based control diet suggesting that time duration of 12 h, 24 h, 36 h and 48 h given for biodegradation of sweet orange peel could not enhance its nutrients. Biodegradation of sweet orange fruit peel for a time frame of 12 h to 48 h yielded a feed ingredient with a relatively high crude fibre, which lowered the growth rate of starter broiler chicks and cannot be used to formulate starter chicks diet at 30% maize replacement.

Keywords: Rumen Content, Biodegradation Duration, Feed Value

1. Introduction

Animal protein is essential in human nutrition because of its biological significance, due to the similarity of its amino acid profile to that of man. The poultry industry is one of the fastest means of providing the much-needed animal protein to reduce shortage in its consumption by man. This requires

optimal management and nutrition to reduce costs and economize poultry meat production to offer high quality products to consumers [1,2]. In developing countries, the increasing cost and decreasing supply of conventional feedstuffs are expected to constrain the future expansion of the livestock industry [3]. The cost of feed alone accounts for about 70-75% of the total cost of broiler production and this

Experimental Diets					
Ingredients (kg)	CD	T ₁ D	T ₂ D	T ₃ D	T ₄ D
<i>Calculated analysis:</i>					
ME (Kcal/kg)	2886.32	2820.60	2798.75	2808.78	2785.81
Crude protein (%)	22.68	22.33	22.41	22.41	22.41
Ether extract (%)	3.88	3.65	3.54	3.53	3.65
Crude fibre (%)	4.24	5.49	5.76	5.69	5.85
Calcium (%)	1.29	1.29	1.29	1.29	1.29
Available P (%)	0.71	0.66	0.66	0.66	0.66
Lysine (%)	1.48	1.44	1.44	1.44	1.44
Methionine (%)	0.74	0.70	0.70	0.70	0.70

*Premix supplies per kg: Vitamin A, 10,000, 000 iu., Vitamin D₃, 2,000,000 mg., Vitamin K₃, 2,000 mg., Vitamin B₁, 3,000 mg., Vitamin B₂, 5,000 mg., Niacin, 45,000 mg., Calcium panthothenate, 10,000 mg., Vitamin B₆, 4,000 mg., Vitamin B₁₂, 20 mg., Choline chloride, 300,000 mg., Folic acid, 1,000 mg., Biotin, 50 mg., Manganese 300,000 mg., Iron 120,000 mg., Zinc, 80,000 mg., Copper, 8,500 mg., Iodine, 1,500 mg., Cabot, 300 mg., Selenium 120 mg., Antioxidant 120,000 mg., BSOP = Biodegraded sweet orange peel.

CD = Control diet containing 0% biodegraded sweet orange peel, T₁D= Diet containing sweet orange peel obtained after 12 h biodegradation, T₂D= containing sweet orange peel obtained after 24 h biodegradation, T₃D= containing sweet orange peel obtained after 36 h biodegradation, T₄D= containing sweet orange peel obtained after 48 h biodegradation.

ME = 37 (%CP) + 81.8 (%EE) + 35.5 (%NFE)-(Pauzenga, 1985).

2.3. Experimental Birds and Management

A total of one hundred and eighty (180) day old broiler chicks (Ross 308) from Sayed Farms were used in the feeding trial. The birds were randomly grouped into five (5) of equal number (36) and similar weight. A group each was assigned also randomly to one of the five (5) experimental diets with three (3) replicates of twelve (12) birds each. The birds were housed in a half-walled deep litter pen with wood shaving as the litter material. The birds were fed *ad-libitum* throughout a 28-day feeding trial and allowed free access to drinking water. Health management protocols involved the administration of infectious bursal disease vaccine (Gumboro) at 10th and 21st days, Newcastle disease vaccine (Lasota) at 14th and 28th days. Anti-stress supplement (vitalyte) was administered at day old, prior to and after each vaccine administration, pre and post weekly weighing of birds. Antibiotics and coccidiostat were also administered routinely each at alternate weeks as prophylactic measures against bacterial infection and coccidiosis.

2.4. Growth Performance Data

Initial and final live weights of the birds were taken at the start and end of the feeding trial using a top-load weighing scale. Weekly body weights were also recorded. Body weight gain (BWG) was determined by weight difference between current and previous weeks, while total weight gain was obtained by the difference between the final and initial live weights. Feed intake was obtained from the amount of feed supplied less the left over. Feed conversion ratio (FCR) was calculated from the ratio of feed consumed to body weight gain (FCR = Feed consumed / BWG) while, protein efficiency ratio (PER) was computed from the ratio of body weight gain to protein consumed (PER = BWG / protein consumed).

2.5. Statistical Analysis

Data generated were subjected to one-way analysis of variance (ANOVA) using [24] and the means of significantly

different ($p < 0.05$) parameters were separated using the Duncan's Multiple Range Test (DMRT) of the same software package.

3. Results and Discussion

The proximate composition and metabolizable energy of sweet orange (*Citrus sinensis*) fruit peel obtained after 12 h, 24 h, 36 h and 48 h biodegradation are presented in Table 2. The crude protein values of the biodegraded sweet orange peel were lower than 8.90% CP in maize, a conventional energy feedstuff [25] used in the formulation of broiler chicken diet. This showed that the biodegraded sweet orange peel used in this study was inferior to maize in crude protein. Higher crude proteins of 10.73% [26] and 8.20% [27] have been reported for non-biodegraded sun dried sweet orange peels. The variation of the CP in this study from those reported by these workers can largely be attributed to the different processing and handling methods. The differences in per cent crude protein present in sweet orange peel reported by the different researchers could be attributed to differences in varieties of orange fruits, season of the year the peels were gathered, processing techniques adopted and the stage of maturity at which the fruits were harvested. The crude fibre (CF) range observed in this study (10.16% to 12.30%) was higher than 2.70% for maize [25] and 7.86% reported for sun dried sweet orange peel [26]. Thus, the biodegradation method used was unable to cause an appreciable reduction in the fibre content of the peel so as to increase its potential use as energy feed ingredient in broiler chicken diet. High fibre level in a feed limits its use in broiler chicken diet. The ether extract (EE) range reported in this study was lower than 4.00% for maize [25]. This may have been responsible for the reduction in carcass visceral fat in a study to evaluate the effect of fermented sweet orange peel on broiler chicken performance, in which lower fat levels in the range of 2.33-2.94% was found in the peel [28] compared to 4.00% in maize; the standard reference energy ingredient. The ash content in the biodegraded orange peel was higher than 2.36% for maize [29] and 6.09% [27] for orange peel

sun dried immediately after collection. Hence, the rumen filtrate may have raised the ash content during biodegradation. Nitrogen free extract (NFE) in this study tended to decrease as the duration of biodegradation of sweet orange peel increased from 0 to 48 h. The possible increase in the growth and proliferation of microbes during biodegradation may be responsible for the reduction of NFE as they will utilize more of the soluble carbohydrates as source of metabolic energy as the duration of the process increased. The metabolisable energy of the biodegraded sweet orange peel had the same trend as the NFE, and lower than 3432 kcal/kg of maize [30]. The result of proximate composition of biodegraded sweet orange peel has elicited the possibility of its being useable in broiler chicken diets if the high fibre content can be reduced by the application of any feed processing method, including microbial technology experimented in this trial.

The quantification of the crude fibre fractions in biodegraded sweet orange (*Citrus sinensis*) fruit peel is presented in Table 3. The duration of 0 to 48 h given for biodegradation of sweet orange peel did not cause significant ($p>0.05$) variation in the acid detergent fibre (ADF), neutral detergent fibre (NDF), acid detergent lignin (ADL),

hemicellulose and cellulose fractions among the treatments. They all appear to decrease marginally as the duration of biodegradation increased. It is this same pattern that was observed in the feed intake of the birds in this study. The level of NDF, ADF, ADL and cellulose in feed stuff are negatively correlated to feed intake of monogastric animals and most especially poultry and pig. This is because of their intrinsic gastrointestinal tract limitation to digest fibrolytic carbohydrates due to the absence of anaerobic bacteria which are carbohydrate fermenters to release energy for their utilisation. Hence, lower amounts of these fibrolytic materials in broiler feed will help to improve feed intake for adequate energy utilisation for productive purposes. The ADF, NDF, and cellulose fractions obtained in this study were comparatively lower than 38.84% to 41.39% ADF, 60.00% to 62.5% NDF, 33.35% to 35.26% cellulose, respectively while, the ADL and hemicellulose were higher than 5.44% to 5.62% ADL and 21.21% to 21.62% hemicellulose, respectively earlier reported [31]. These variations might be due to the differences in the production of fibrolytic enzymes according to the substrates and microorganisms during fermentation [32].

Table 2. Proximate Composition and Metabolizable Energy of Biodegraded Sweet Orange Fruit Peel Meal (% DM).

Biodegraded Sweet Orange Peel Meal				
Parameters	BSOP ₁₂	BSOP ₂₄	BSOP ₃₆	BSOP ₄₈
Dry matter	90.43	90.38	89.82	90.50
Crude protein	6.78	7.26	7.30	7.25
Crude fibre	10.16	11.76	11.38	12.30
Ether extract	2.63	1.97	1.88	2.65
Ash	7.97	9.23	7.79	11.76
Nitrogen free extract	72.46	69.78	71.65	66.04
Metabolizable energy (Kcal/kg)	3037.97	2906.96	2967.46	2829.44

BSOP₁₂: Biodegraded sweet orange peels obtained after 12 h fermentation.

BSOP₂₄: Biodegraded sweet orange peels obtained after 24 h fermentation.

BSOP₃₆: Biodegraded sweet orange peels obtained after 36 h fermentation.

BSOP₄₈: Biodegraded sweet orange peels obtained after 48 h fermentation.

Metabolizable energy = 37 (%CP) + 81.8 (%EE) + 35.5 (%NFE) (Pauzenga, 1985).

Table 3. Fibre Fractions in Biodegraded Sweet Orange Fruit Peel Meal (% DM).

Biodegraded Sweet Orange Peel Meal				
Fibre fractions	BSOP ₁₂	BSOP ₂₄	BSOP ₃₆	BSOP ₄₈
ADF	22.30	21.20	22.50	19.50
NDF	56.10	54.40	56.70	52.30
ADL	8.40	7.60	8.10	6.80
Hemicellulose	33.80	33.20	34.20	32.80
Cellulose	13.90	13.60	14.40	12.90

ADF: Acid detergent fibre, NDF: Neutral detergent fibre, ADL: Acid detergent lignin.

BSOP₁₂: Sweet orange peels obtained after 12 h biodegradation.

BSOP₂₄: Sweet orange peels obtained after 24 h biodegradation.

BSOP₃₆: Sweet orange peels obtained after 36 h biodegradation.

BSOP₄₈: Sweet orange peels obtained after 48 h biodegradation.

The performance of starter broiler chicks in the feeding trial is presented in Table 4. The mean final live body weight of starter broiler chicks showed that the birds in the control group (CD) were significantly ($p<0.05$) heavier than the birds in the biodegraded sweet orange peel meal (BSOP) based diets. This is possibly a direct consequence of the nature of the

experimental diets because, the biodegraded sweet orange peel used as a replacement for maize at 30% level had a lower feed value due to its high fibre content. This conferred lower nutrient status on the BSOP based diets hence, the significant ($p<0.05$) variation in the body weight between the chicks in the control and the test diet groups. The birds fed the BSOP

based diets had similar final live body weight which showed that the time allowed for biodegradation of sweet orange peel which ranged from 12 h to 48 h did not enhance the nutrients in it. The broiler starter live weight obtained agrees with 491.00 g to 831.67 g when sweet orange peel meal was incorporated into broiler chick diets [15], and it has been earlier reported that broiler chickens on control diets were heavier than those on sweet orange peel meal-based diets [28, 33, 34]. The total weight gain, daily weight gain, feed conversion ratio and protein efficiency ratio also differed significantly ($p < 0.05$) and followed the same trend as the final weight. Daily feed intake and protein intake were also affected significantly ($p < 0.05$) by the experimental diets with a direct relationship between these performance indicators. The daily feed intake obtained in this study was observed to be comparable to 37.28 g to 44.64 g obtained when exogenous enzyme was added to treat sweet orange peel used as

replacement for maize [27] but, lower than 52.68 g to 62.56 g reported by [26] when sun-dried sweet orange peel meal was used in broiler starter diets. Birds fed diets containing biodegraded sweet orange peel meal had similar daily feed intake, feed conversion ratio and daily protein intake. This suggests that biodegradation of sweet orange peel using rumen filtrate from cattle for durations of 12 h to 48 h did not upgrade the feed quality of sweet orange peel. Feed conversion ratio (FCR) obtained was similar to 1.54-1.70 when exogenous enzyme was used to treat sweet orange peel [27]. Feed conversion rate is usually low for young animals because of their fast relative growth, and increases for older animals when relative growth tends to flatten out and feed intake is high. Mortality occurred in all the dietary groups with a pattern difficult to link to the experimental diets. An earlier report on sweet orange peel meal showed 0% mortality even at higher percentages of maize replacement with sweet orange peel [35].

Table 4. Performance of Starter Broiler Chicks fed Diets Containing Biodegraded Sweet Orange Peel Meal.

Experimental Diets						
Performance Indices	CD	T ₁ D	T ₂ D	T ₃ D	T ₄ D	SEM
Initial live weight (g/bird)	36.11	35.42	36.11	35.42	36.11	0.54 ^{ns}
Final live weight (g/bird)	811.43 ^a	617.20 ^b	590.97 ^b	576.14 ^b	583.83 ^b	24.73
Total weight gain (g/bird)	775.31 ^a	581.78 ^b	554.86 ^b	540.72 ^b	547.72 ^b	24.37
Daily weight gain (g/bird/day)	27.69 ^a	20.73 ^b	19.82 ^b	19.31 ^b	19.56 ^b	0.87
Daily feed intake (g/bird/day)	43.06 ^a	40.62 ^{ab}	38.67 ^{ab}	38.47 ^b	38.21 ^b	1.35
Feed conversion ratio	1.56 ^b	1.95 ^a	1.95 ^a	1.99 ^a	1.95 ^a	0.03
Daily protein intake (g/bird/day)	9.77 ^a	9.07 ^{ab}	8.67 ^b	8.62 ^b	8.56 ^b	0.30
Protein efficiency ratio	2.83 ^a	2.29 ^b	2.29 ^b	2.24 ^b	2.29 ^b	0.03
Mortality (%)	2.78 ^{ab}	11.11 ^a	0.00 ^b	5.55 ^{ab}	2.78 ^{ab}	2.48

^{a, b}Means in the same row with different superscripts are highly significantly different ($p < 0.05$), SEM = Standard error of mean, ^{ns}Not significantly different ($p > 0.05$).

CD = Control diet containing 0% biodegraded sweet orange peel meal, T₁D= Diet containing sweet orange peel obtained after 12 h biodegradation, T₂D= containing sweet orange peel obtained after 24 h biodegradation, T₃D= containing sweet orange peel obtained after 36 h biodegradation, T₄D= containing sweet orange peel obtained after 48 h biodegradation.

4. Conclusion

The birds fed the biodegraded sweet orange peel (BSOP) based diets had similar final live body weight, body weight gain, feed conversion ratio and protein efficiency ratio. The growth indices for the birds in the BSOP dietary treatments were inferior to the birds on the maize based control diet suggesting that the duration of 12 h, 24 h, 36 h and 48 h given for biodegradation of sweet orange peel did not enhance its nutrients. Biodegradation of sweet orange fruit peel for a time frame of 12 h to 48 h yielded a feed ingredient with a relatively high crude fibre, which lowered the growth rate of starter broiler chicks, and hence cannot be used to formulate starter broiler chicks diet at 30% maize replacement.

References

- [1] Pope, T. and Emmert, J. L. (2001). Phase-Feeding Supports Maximum Growth Performance of Broiler Chicks from Forty-three to Seventy-one Days of age. *Journal of Poultry Science*, 80: 345-352.
- [2] Laudadio, V., Tufarelli, V., Dario, M., D'Emillo, F. P. and Vicenti, A. (2009). Growth performance and carcass characteristics of female turkeys as affected by feeding programs. *Journal of Poultry Science*, 88: 805-810.
- [3] FAO (2012). FAOSTAT. Food and Agriculture Organization of the United Nations.
- [4] Jurgens, M. H., Lee, I and Chibam, C. (2009). Poultry Nutrition and Feeding. *Animal Nutrition Handbook*. 2002; 316. Available: <http://www.ag.auburn.edu/~chibale/an12poultryfeeding.pdf>
- [5] Sobamiwa, O. and Akinwale, T. O. (1999). Replacement value of cocoa husk meal for maize diets in growing pullets. *Tropical Journal of Animal Science*, 1: 111-116.
- [6] Tuleun, C. D., Njike, M. C., Ikurior, S. A. and Ehiobu, N. G. (2005). Laying performance and egg quality of hens fed cassava root meal/brewer's yeast slurry-based diets. *Production Agriculture and Technology*, 1: 148-152.
- [7] Oluremi, O. I. A., Ojighen, V. O and Ejembi, E. H. (2006). The nutritive potentials of sweet orange (*Citrus sinensis*) in Rind broiler production. *International Journal of Poultry Science*, 5: 613-617.

- [8] Oluremi, O. I. A., Gabriel, O. S., Ipirakwagh, E. N., Ikwue, C. O. and Afolabi, E. T. (2018). Performance and blood profile of rabbits fed biodegraded sweet orange (*Citrus sinensis*) peel-based diet. *Nigerian Journal of Animal Science*, 20 (3): 287-297.
- [9] Famurewa, J. A. V. and Olarewaju, A. (2013). Investigating the potentials dried palm oil mill effluents from pressing and water displacement methods for animal feed. *Asian Journal of Natural and Applied Science*, 2: 58-68.
- [10] Orayaga, K. T., Oluremi, O. I. A., Tuleun, C. D. and Carew, S. N (2015). The feed value of composite mango (*Mangifera indica*) fruit reject meal in the finisher broiler chickens' nutrition. *African Journal of Food Science and Technology*, 6 (6): 177-184.
- [11] Orayaga, K. T. (2016). Effect of composite mango (*Mangifera indica*) fruit rejects meal on growth performance, digestibility and economics of production of Rabbits. *Nigerian Journal of Animal Science*, 1: 65-75.
- [12] Olanrewaju, R. I. (2020). Orange Production in Nigeria. Sence Agriculture, Crop Production <https://www.agriculturenigeria.com/production/orange-production/>
- [13] Knoema.com (2018). World atlas: Nigeria-Citrus fruit production quantity <https://knoema.com/atlas/Nigeria/topics/Agriculture/Crops-Production-Quantity-tonnes/Citrus-fruit-production>.
- [14] Ipinjolu, J. K. (2000). Performance of juvenile orange koi carp (*Cyprinus carpio L.*) fed diets supplemented with sweet orange peel meal: body composition, nutrition, utilization and skin pigmentation. *Sokoto Journal of Veterinary Science*, 2000: 228-229.
- [15] Oluremi, O. I. A., Okafor, F. N., Adenkola, A. Y. and Orayaga, K. T (2010). Effect of fermentation of sweet orange (*Citrus sinensis*) fruit peels on its phytonutrients and performance of broiler starter. *International Journal of Poultry Science*, 9: 546-549.
- [16] Agu, P. N. (2006). Nutritive value of sweet orange (*Citrus sinensis*) peel as a feed resource in broiler production. *Unpublished M. Sc Thesis*, Department of Animal Production, University of Agriculture, Makurdi, Nigeria.
- [17] Worldometer (2020) Nigeria Population. <https://www.worldometers.info/world-population/nigeria-population/>
- [18] Wikipedia (2020). Makurdi: Location in Nigeria. <https://en.wikipedia.org/wiki/Makurdi>.
- [19] Custom Weather (2020). Climate and Weather Averages in Makurdi, Nigeria: Weather Reports Collected between 2005-2015. <https://www.timeanddate.com/weather/nigeria/makurdi/climate>.
- [20] TAC. (2011). Makurdi Weather Element Records. Meteorological station Nigeria Air Force, Tactical Air Command, Makurdi, Nigeria.
- [21] A. O. A. C. (2002). Association of Official Analytical Chemists. 17th Edition. William Press. Richard Virginia, USA.
- [22] Goering, H. K. and Van Soest, P. J. (1970). Forage fibre analysis. USDA Agriculture Handbook No. 379. USDA-ARS, Washington, DC.
- [23] Pauzenga, U. (1985). Feeding Parent Stock. *Journal of Zootechnica International*, 19: 22-24.
- [24] SPSS.com (2012). IBM® SPSS®. Advantage for Micosoft® IBM. Corporation 2012, IBM corporation route, 100 Somers, N4.10589.
- [25] Aduku, A. O. (2004). Animal Nutrition in the Tropics. Feeds and Feeding, Pasture management, Monogastric and Ruminant Nutrition, 1st Edition University Press, ABU, Zaria, Nigeria. Pp. 128-134.
- [26] Agu, P. N., Oluremi, O. I. A. and Tuleun, C. D. (2010). Nutritional evaluation of sweet orange (*Citrus sinensis*) fruit peel as feed resources in broiler production. *International Journal of Poultry Science*, 9 (7): 684-688.
- [27] Sunmola, T. A., Tuleun, C. D. and Oluremi, O. I. A. (2018). Performance characteristics of starter broiler chicks fed dietary sun-dried sweet orange peel meal (SOPM) with and without polyzyme®. *Scientific Research Journal (SCIRJ)*. 6 (8): 89-97.
- [28] Akpe, M. E., Oluremi, O. I. A. and Tuleun, C. D. (2019). Haematological and Serum Biochemical Indices of Broiler Chickens Fed Diets Containing Graded levels of Biodegraded Sweet Orange (*Citrus sinensis*) Peel. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 12 (8), 2: 54-59.
- [29] Ape, D. I., Nwogu, N. A., Uwakwe, E. I. and Ikedinobi, C. S. (2016). Comparative Proximate Analysis of Maize and Sorghum Bought from Ogbete Main Market of Enugu State, Nigeria. *Greener Journal of Agricultural Science*, 6 (9): 272-275.
- [30] Aduku, A. O. and Olukosi, J. O. (2000). Animal products processing and handling in the tropics. In: Living book series. 3rd edition, Abuja. G. U. Publications. Pp. 32.
- [31] Oluremi, O. I. A., Ngi, J. and Andrew, I. A (2007). Phytonutrients in *Citrus* fruit peel meal and nutritional implications for livestock production. *Livestock Research for Rural Development*, 9: 89.
- [32] Shi, C., He, J., Yu, J., Yu, B., Huang, Z., Mao, X., Zheng, P. and Chen, P. (2015). Solid state fermentation of rapeseed cake with *Aspergillus niger* for degrading glucosinolates and upgrading nutritional value. *Journal of Animal Science and Biotechnology*, 6: 13-19.
- [33] Oluremi O. I. A., Ahile A. A. and Jande T. F (2017). Effect of feeding graded levels of fermented sweet orange (*Citrus sinensis*) fruit peel meal on the growth and nutrient digestibility of broiler chicken. *International Journal of Environment, Agriculture and Biotechnology*, 2: 3119-3123.
- [34] Nwobodo, E. A., Oluremi, O. I. A., Tuleun, C. D. and Kaankuka, F. G. (2020). The effect of Biodegraded Sweet Orange (*Citrus Sinensis*) Fruit Peel on the Growth and Economic Performance of Starter Broiler Chicks. *Journal of Animal Husbandry and Dairy Science*, 4 (1): 23-29.
- [35] Oluremi, O. I. A., Mou, P. M. and Adenkola, A. Y. (2008). Effect of sweet orange (*Citrus sinensis*) fruit peels on its maize replacement value in broiler diet. *Livestock Research for rural Development*, 20: 246-298.