

Blood and Gut Microbiota Profiles of Broiler Chickens Fed on Diet Supplemented with Graded Levels of Neem (*Azadirachta indica*) Oil

Mafouo Sonhafouo Vanessa*, Kana Jean Raphaël, Nguepi Ndogmo Kissel

Animal Nutrition and Production Research Unit, Department of Animal Science, University of Dschang, Dschang, Cameroon

Email address:

vanessa.sonhafouo@yahoo.fr (M. S. Vanessa)

*Corresponding author

To cite this article:

Mafouo Sonhafouo Vanessa, Kana Jean Raphaël, Nguepi Ndogmo Kissel. Blood and Gut Microbiota Profiles of Broiler Chickens Fed on Diet Supplemented with Graded Levels of Neem (*Azadirachta indica*) Oil. *Animal and Veterinary Sciences*.

Vol. 7, No. 3, 2019, pp. 78-82. doi: 10.11648/j.avs.20190703.12

Received: April 17, 2019; **Accepted:** June 5, 2019; **Published:** June 26, 2019

Abstract: The present study was assigned to evaluate the Hematological and gut microbiota profiles of broiler chickens fed on diet supplemented with graded levels of neem oil. A total of 400 day-old chicks of Cobb 500 strain were randomly assigned to 5 treatments groups of 80 birds each with 5 replicates. The experimental rations consisted of a control diet without supplement (R0-), a positive control diet containing 1g of antibiotic/kg (R0+) and three other diets supplemented respectively with 15, 20 and 25 g of neem oil / kg of feed. Feeding broiler with neem oil had no significant ($p < 0.05$) effect on hemoglobin (Hb), Packed cell volume (PCV), Red blood cell (RBC), total leucocytes, white blood cells (WBC), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC) and lymphocytes counts. With 25 g of oil / kg no trace of *salmonella* has found in the digestive tract. While, *shigellae* count was significantly lower with 20 g of neem / kg feed compared to the negative control diet. It was concluded that up to 20 g of neem could be included in a kg of broiler chickens diet without deleterious effect on their blood constituents and with the benefit of reducing possible risks of infection from pathogenic bacteria.

Keywords: Neem Oil, Hematology, Bacterial Counts, Broiler Chickens

1. Introduction

The use of synthetic antibiotics as growth promoters has high cost implications and adverse side effect on bird's health, prolonged withdrawal period and risk of accumulation in tissues and egg which could have harmful effects on human health [1]. As a result, consumers of poultry products are demanding for drug residues free meat. This has triggered the search for alternatives means to produce birds at reduced cost using natural growth and health promoters. Studies of neem (*Azadirachta indica*) indicate an increasing interest in the plant owing to its versatile application and promising uses.

Neem (*Azadirachta indica*), popularly known as Indian neem (margosa) or Indian lilac of the family *Maliaceae*, is one of such non-conventional and available ingredients sources in the tropics with great potential. It is a tropical tree

plant which is widely distributed in Africa and available all year round [2, 3]. The tree is well adapted to the climatic and soil conditions in the tropical rainforest regions, all the way to the Sahel savannah part of Cameroon. The leaves are very bitter to taste, and possess a garlic-like smell. The plant is popular because it is readily available, cheap, non-toxic to animals and humans and efficacious against malaria [3].

A. indica seed oil contains triterpenoid compounds such as azadirachtin, gedunine, nimbine and nimbidine which have antibacterial and antifungal properties [4, 5]. The blend of these compounds in poultry feed can produce additive or synergistic effects on production performance [5]. The antimicrobial activity of extracts of neem leaves against such micro-organisms as *Staphylococcus* spp, *Streptococcus* spp, *Pseudomonas* spp and *Escherichia coli*, and some fungal strains have been reported [3, 5]. Studies on the effects of neem leaves on poultry production especially of broilers and

laying hens also exist [6, 7]. Antimicrobial studies on the effects of neem leaves and their extracts on cultured microorganisms *in vitro* have also been carried out [3].

However, there is no information on the performance, the health and the physiological conditions of animal fed on neem oil. The present study was therefore undertaken to investigate the effects of graded levels of neem oil on the hematological profiles and gut microbial counts of broiler chickens.

2. Materials and Methods

2.1. Study Area

This study was carried out at the Poultry Unit of the Teaching and Research Farm of the Faculty of Agronomy and Agricultural Sciences of the University of Dschang, Cameroon. Dschang is located in the West region of Cameroon at 05° 26' N and 10° 26' E. The area experiences a wet season from March to November and a hot dry season for the rest of the year. Maximum ambient temperature is around 21°C, while an average annual rainfall of 2000 mm is prevalent.

2.2. Processing of Neem Oil and Experimental Rations

Fresh neem fruits were collected at Garoua in the Northern Region of Cameroon between September and October 2017. They were thoroughly washed with running tap water and separated from the sheet manually. The seeds were oven dried at 50°C until constant moisture content was achieved. The dried seeds were ground in a mill for size reduction and the extraction of oil was carried out by kneading the paste. The phytochemical analyses of neem oil carried out as described by Talukdar *et al.* [8] revealed that alkaloids, flavonoids, terpenoids, phenols, steroids and tannins were presents.

Experimental diet consisted of control ration without no supplement (R₀⁻), positive control supplemented with 1 g of antibiotic /kg of feed (R₀⁺) and 3 others rations supplemented with 15 g (R₁₅), 20 g (R₂₀) and 25 g (R₂₅) of neem oil/kg of feed. The antibiotic used in positive control ration was Doxycycline[®]. The proximate composition of the basal diet is summarized in Table 1.

Table 1. Composition of the basal diet.

Ingredients (%) Starter		Finisher
Maize	59	65
Remoulage	5	5
Cotton seed cake (50%)	6	4
Soybean meal (49%)	20	15
Fish meal (60%)	4	5
Shellfish	0.75	1
Bone meal	0.25	0
Premix 5%*	5	5
Total	100	100
Analysed composition		
Metabolizable energy (kcal/kg)	2951.91	3006.85
Crude protein (%)	22.53	20.38
Calculated composition		

Ingredients (%) Starter		Finisher
Energy/Protéin	130.99	147.54
Lysine (%)	1.32	1.19
Methionine (%)	0.46	0.45
Calcium (%)	1.07	1.15
Non-phytate phosphorus (%)	0.5	0.48
Calcium/phosphore	2.16	2.41
Crude fibers (%)	4.96	4.94
Price/kg (FCFA)	273.59	147.54

* Premix: Mineral Nitrogen and Vitamin Complex: Crude protein = 40%, Calcium = 8%, Phosphorous = 2.05%, Lysine = 3.3%, Methionine = 2.40%, ME = 2078 Kcal / kg, Vit A: 3,000,000UI, Vit D 3: 600,000UI, Vit E: 4,000mg, Vit K: 500mg, Vit B1: 200mg, VitB2: 1000mg, Vit B6: 400mg, Vit B12: 4mg, Iron: 8000mg, Cu: 2000mg, Zn: 10,000mg, Se: 20mg, Mn: 14000mg

2.3. Experimental Birds

A total of 400-day-old Cobb 500 strain chicks with an average body weight of 38.53 g obtained from a local hatchery has provided with *ad libitum* access to feed and water. Birds were given vaccines in drinking water against Newcastle disease and Infectious Bronchitis on the 8th day with a booster dose on the 23th day of age, and against Gumboro disease on the 10th day of age. Coccidiosis prevention was done using Vetacox[®] for 3 consecutive days per week from the 2nd to the 6th week of age. Birds were administered commercial antistress (Amintotal[®]: LAPROVET, Tours Cedex 2, France) in drinking water during the first 3 days upon arrival, after each vaccination, weighing session and transfer from brooding house to finishing house.

The birds were weighed at the beginning of the experiment and at weekly intervals thereafter. The birds were randomly assigned to the five treatment dietary groups (R0+, R0-, R15, R20 and R25) of 80 birds each. Each treatment was further divided into five (5) replicates of sixteen (16) birds and places in pens in line with the design of the experiment.

2.4. Hematological Parameters

At 7 weeks, ten birds (05 males and 05 females) from each treatment were randomly selected for bleeding. 3mL of the blood was collected from each bird into a tube containing EDTA as anticoagulant, and shaken gently to prevent coagulation. Blood samples were immediately analyse for hematological parameters using routine laboratory procedures as described by Schalm *et al.* [9] Saror and Schillhorn van Veen [10], Friedman and Young [11], Iyayi and Tewe [12]. The hemoglobin (Hb) content was determined with a digital photo colorimeter (Model 312E by Digital Photo Instruments, Germany) at a wavelength of 625nm and expressed in gram (g) units. Packed cell volume (PCV) was determined through the Winthrose microhaematocrit technique, and expressed in percentages (%). Red blood cell (RBC) counts were obtained with a Coulter Electronic counater (Model ZF by Coulter Electronic Ltd. London), and the values expressed in millions per microlitre (μL) of blood (x10⁶/μL). The white blood cells (WBC) were counted with an improved Neubauer

hemocytometer and expressed in thousands per microlitre (μL). Other hematological parameters computed or evaluated included mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), and mean corpuscular haemoglobin concentration (MCHC) and lymphocytes counts.

2.5. Gut Microbiota Count

Sterile forceps were used to collect the faecal samples from each treatment group from the dropping tray into anal swab. Microbial count and identification were done using routine laboratory procedures outlined by Duraipandiyar *et al.* [5], Tuhin *et al.* [13] and Valarmathy *et al.* [14]. The analysis included preparation of culture media, microscopic and macroscopic examination of the faecal samples, and growth/population counts. Total Bacteria Counts were then determined. All glassware used were thoroughly washed with detergent, rinsed with distilled water, wrapped in aluminum foil and sterilized in a hot air oven at 125°C for 1 hour.

2.6. Data Analysis

Data collected were subjected to one-way analysis of variance (ANOVA) according to the procedure of Steel and Torrie [15]. Where ANOVA indicated significant different treatment means, they were separated using the Duncan's New Multiple Range Test at 5% [16].

3. Results

The summary of the effects of inclusion levels of neem on the hematological parameters of the broiler chickens is presented in Table 2. The analysis of variance revealed no significant ($p > 0.05$) difference between treatment groups regardless of the characteristics evaluated. However, although not significant, the number of white blood cells, granulocytes, blood platelet count (PLT) and platelets (PCT) tend to increase with the increasing level of neem oil in the ration.

Table 2. Effects of graded level of neem oil in the ration on the hematological parameters of broiler chickens.

Hematological characteristics	Control		Neem oil (g / kg of food)			p
	R ₀ ⁻	R ₀ ⁺	15	20	25	
WBC ($10^3 \mu\text{l}$)	9.29 \pm 5.45	6.5 \pm 1.31	7.83 \pm 1.57	8.50 \pm 2.64	8.94 \pm 5.13	0.644
LYM ($10^3 \mu\text{l}$)	4.80 \pm 1.53	2.98 \pm 0.72	2.87 \pm 0.76	2.85 \pm 0.93	2.96 \pm 1.75	0.409
GRAN ($10^3 \mu\text{l}$)	3.36 \pm 1.54	2.84 \pm 1.31	3.84 \pm 0.68	4.57 \pm 1.56	4.80 \pm 1.90	0.394
RBC ($10^6 \mu\text{l}$)	3.66 \pm 0.49	4.10 \pm 0.41	4.12 \pm 0.49	4.94 \pm 1.97	4.15 \pm 0.32	0.185
Hb (g/dl)	12.10 \pm 2.22	13.44 \pm 2.44	13.23 \pm 1.63	16.35 \pm 6.16	13.96 \pm 1.76	0.190
HCT (%)	30.36 \pm 4.97	33.17 \pm 5.79	33.10 \pm 5.28	40.43 \pm 5.78	34.95 \pm 4.33	0.245
MCV (fL)	83.00 \pm 5.99	80.58 \pm 9.40	80.78 \pm 11.53	82.31 \pm 12.37	84.46 \pm 8.94	0.939
MCH (pg)	32.89 \pm 2.30	32.54 \pm 4.07	32.30 \pm 3.88	33.31 \pm 4.27	33.64 \pm 3.70	0.957
MCHC (g/dl)	39.84 \pm 3.25	40.44 \pm 1.46	40.21 \pm 3.09	40.65 \pm 1.42	39.91 \pm 1.94	0.955
PLT ($10^3 \mu\text{l}$)	24143 \pm 128.89	176 \pm 76.25	198.86 \pm 48.55	210.50 \pm 102.72	294.25 \pm 145.76	0.267
PCT (%)	0.30 \pm 0.15	0.22 \pm 0.09	0.25 \pm 0.05	0.26 \pm 0.14	0.38 \pm 0.19	0.241

R₀⁻: 0g of neem oil / kg of food; R₀⁺: R₀⁻ + 1g of Doxycycline / kg of food; p: Probability WBC: white blood cells; Lym: lymphocytes; Gran: granulocytes; RBC: red blood cells; Hb: hemoglobin; HCT: hematocrit; PLT: platelet count; PCT: plaquetocrite; MCH: mean corpuscular hemoglobin; MCHC: Mean corpuscular concentration of hemoglobin.

The bacteria count was significantly ($p < 0.05$) affected by the increasing level of neem oil (Table 3). The animals fed on diet supplemented with 25 g of neem oil / kg were not infested with salmonella, while the number of *Salmonella* colonies was significantly higher ($p < 0.05$) in chickens fed on the control diet without supplement. The supplementation of the rations with neem oil whatever the doses stimulated the multiplication ($p < 0.05$) of *Lactobacilli* compared to the

negative control diet. The number of *shigellae* recorded in animals supplemented with 20 g of neem oil / kg of feed was significantly lower compared to the number induced by the control diet without supplement. The *Escherichia coli* counts recorded in chickens fed on different levels of neem oil was statistically ($p < 0.05$) higher compared to the chickens fed antibiotic medicated diet (Table 3).

Table 3. Variation of gut bacteria (CFU / swab) of the broiler fed on graded levels of neem oil supplemented diet.

Number of bacteria colony (Log ₁₀ CFU / swab)	Control		Neem oil (g / kg of food)			p
	R ₀ ⁻	R ₀ ⁺	15	20	25	
<i>Escherichia coli</i>	4.65 \pm 0.46 ^a	3.80 \pm 0.49 ^b	4.62 \pm 0.70 ^a	4.98 \pm 0.60 ^a	4.66 \pm 0.19 ^a	0.061
<i>Salmonella</i>	1.53 \pm 0.70 ^a	0.62 \pm 0.13 ^a	1.51 \pm 0.17 ^a	0.85 \pm 0.19 ^a	0.00 \pm 0.00 ^b	0.055
<i>Shigella</i>	5.27 \pm 0.55 ^a	3.28 \pm 1.32 ^{ab}	4.15 \pm 1.77 ^{ab}	1.19 \pm 0.38 ^b	4.01 \pm 1.69 ^{ab}	0.051
<i>Lactobacilli</i>	4.79 \pm 0.39 ^b	5.44 \pm 0.40 ^{ab}	5.70 \pm 0.34 ^a	5.67 \pm 0.82 ^a	5.78 \pm 0.44 ^a	0.059

a, b: averages with the same letter on the same line are not significantly different ($P > 0.05$). R₀⁻: 0g of neem oil / kg of food; R₀⁺: R₀⁻ + 1g of Doxycycline / kg of food; p: Probability

Table 4 shows that body weight and weight gain are negatively correlated with the number of bacteria regardless

of the rate of oil and the type of bacteria present in the digestive tract. Moreover, regardless of the oil content, the

feed conversion ratio is positively correlated to *Salmonella* and *Shigella* counts. At the incorporation rate of 25 g of neem oil/kg of feed, the feed conversion ratio is significantly, strongly and positively correlated with the number of

Escherichia coli. With 20 g of neem oil/kg, the number of *Salmonella* and *Shigella* is positively and significantly ($p < 0.05$) correlated to the the feed conversion ratio.

Table 4. Correlation coefficients between growth parameters, bacterial counts and incorporation rate of neem oil.

Incorporation rate of neem oil (g/kg of feed)	Growth parameters	Bacteria			
		<i>Escherichia coli</i>	<i>Salmonella sp</i>	<i>Shigella sp</i>	<i>Lactobacillus sp</i>
0	FI	-0.310	0.354	-0.310	0.070
	LW	0.078	0.221	-0.105	-0.432
	WG	-0.103	0.297	-0.014	-0.566
15	FC	-0.007	-0.142	-0.049	0.437
	FI	0.262	0.393	0.709	0.464
	LW	-0.633	-0.490	-0.477	-0.440
20	WG	-0.506	-0.597	-0.622	-0.575
	FC	0.502	0.575	0.614	0.556
	FI	-0.439	0.705	0.705	-0.005
25	LW	0.378	-0.859	-0.859	-0.848
	WG	0.362	-0.755	-0.755	-0.865
	FC	-0.446	0.908*	0.908*	0.806
25	FI	0.106	0.000 ^a	0.144	0.048
	LW	-0.989**	0.000 ^a	-0.861	0.447
	WG	-0.975**	0.000 ^a	-0.856	0.421
	FC	0.915*	0.000 ^a	0.758	-0.484

*Significant at 0.05 ** Significant at 0.01

FI: feed intake; LW: live weight; WG: weight gain; FC: feed conversion

4. Discussion

Blood parameters are considered as the main pathological [17] and nutritional [18] physiological indices for assessing the state of an organism. Any change in the constituent elements of the blood relative to the normal values is an important index for the interpretation of the physiological or metabolic state of the animal but also and especially the quality of the food [19]. Decrease in red blood cells is usually associated with poor quality foods. Hemoglobin determines an animal's ability to withstand a certain level of respiratory stress and hematocrit is the proportion of blood containing red blood cells [20]. When their values are high, this characterizes a polycythemia and when they are weak, it indicate anemia [21]. In the present study, the increasing rate of neem oil tends to increase red blood cell, hematocrit and hemoglobin counts compared to the negative and positive controls groups. This result is in agreement with the findings of Vivian *et al.* [22] who recorded an increase in the number of red blood cells and hemoglobin in chickens receiving aqueous extracts of ginger. Although not significant, the levels of white blood cells, lymphocytes and granulocytes tend to increase with the increasing level of this phytobiotic. This result corroborates the findings of Odoh and Lawrence [23] who reported that by incorporating 10 and 15% neem leaf powder into the broiler food this increased the content of white blood cells and lymphocytes. The increase in white blood cells could be explained by the presence of the active compounds like triterpenoids and phenols in neem oil which might play a very decisive role in stimulating the body's immune system. In this study, the values of hematological parameters observed with the incorporation of neem oil in the

ration remained within the range of physiological reference values recommended by Almosni-Le Sueur *et al.* [24].

The *lactobacilli* count was significantly higher with rations supplemented with neem oil irrespective of the dose. With 25 g of oil / kg no trace of *salmonella* was observed in digestive tract; while, the number of *shigellae* recorded was significantly lower with the ration supplemented with 20 g of neem / kg oil compared to the negative control diet. These results are in agreement with the finding of Divya *et al.* [25] who reported the antibacterial effect of neem oil on several bacteria isolated from a drug-resistant human. *In vitro* studies conducted by Abalaka *et al.* [26] revealed that neem oil at 50 mg has inhibitory effects on the growth of *Pseudomonas aeruginosa*, *Klebsiella ozanae*, *Staphylococcus aureus* and *Escherichia coli*.

5. Conclusion

This study revealed that administration of neem oil to broiler chickens through feed has no significant effect on hematological parameters. However, whatever the incorporation rate, neem oil can be used as feed additive to balance gut microbiota thus ensuring a good health of the digestive tract which is a key factor of a better growth in animal.

References

- [1] Jawad Z., Younus M., Rehman M. U., Munir R., Maqbool A., Shahzad W., Masood S. and Muhammad A., 2014. Effect of *Azadirachta indica* on the Hepatorenal functions in broilers chickens. *The Journal of Animal and Plant Sciences*, 24 (4): 1012-1018.

- [2] Ogbuewu I. P., Odoemenam V. U., Obikaonu H. O., Opara M. N., Emenalom O. O., Uchegbu M. C., Okoli I. C., Esonu B. O. and Iloje M. U., 2011. The Growing Importance of Neem (*Azadirachta indica* A. Juss) in Agriculture, Industry, Medicine and Environment: A Review. *Research Journal of Medicinal Plants*, 5 (3): 230–245.
- [3] Koona S. and Budida S., 2011. Antibacterial Potential of the Extracts of the Leaves of *Azadirachta indica* Linn. *Notulae Scientia Biologicae*, 3 (1): 65–69.
- [4] Makeri H. K., Maikai V. A, and Nok J. A., 2007. Effect of Topical Application of Neem Seed (*Azadirachta indica*) Extract on Sheep Infested with *Amblyomma variegatum*. *African Journal of Biotechnology*, 6 (20): 2324–2327.
- [5] Valarmathy K., Gokulakrishnan Salma K. M. and Kusum D. P., 2010. A Study of Antimicrobial Activity of Ethanolic Extracts of Various Plant Leaves Against Selected Microbial Species. *International Journal of Pharmacology Sciences and Research*, 1 (8): 293–295.
- [6] Esonu B. O. and Iloje M. U., 2011. The Growing Importance of Neem (*Azadirachta indica* A. Juss) in Agriculture, Industry, Medicine and Environment: A Review. *Research Journal of Medicinal Plants*, 5 (3):230–245.
- [7] Obun C. O., Ukim C. I., Olatunji E. A. and Kehinde A. S., 2013. Health and carcass implications of dietary inclusion of graded level of sun cured Neem (*Azadirachta indica*, A. Jus) leaf meal for broilers. *Greener Journal of Agricultural Science*, 3 (1): 48-54.
- [8] Talukdar, A. D., Choudhary, M. D., Chakraborty, M., Dutta BK., 2010. Phytochemical screening and TLC profiling of plant extracts *Cyathea gigantea* (Wall. Ex. Hook.) Haltt and *Cyathea brunoniana* Wall. Ex. Hook. (Cl. & Bak.). *Journal of Science and Technology: Biological and Environmental Sciences* 5: 70-74.
- [9] Schalm O. W., Jain N. C. and Carol, E. J., 1975. Veterinary Hematology. 3rd Edition. *Lea and Febiger, Philadelphia*. Pp 51-81.
- [10] Saror D. I. and Schillhorn van Veen T. W., 1977. Haematological Values of Uda and Yankasa Sheep in the Northern Guinea Savanna of Nigeria. *Tropical Animal Health and Production*, 9 (4): 245-248.
- [11] Friedman R. B. and Young D. S., 1997. Effect of Disease on Clinical Laboratory Tests. 3rd Edition. *AACC Press, Washington DC*.
- [12] Iyayi E. A. and Tewe O. O., 1998. Serum Total Protein, Urea Creatinine Levels as Indices of Quality of Cassava Diets for Pigs. *Tropical Veterinary*, 16: 59-67.
- [13] Durairandiyan V., Muniappan A. and Savarimthu I., 2006. Antimicrobial Activity of some Ethnomedicinal Plants Used by Paliyer Tribe from Tamil Nadu, India. *Entomology Research Institute Loyola College Chennai*. 60034, India.
- [14] Duncan D. B., 1955. New Multiple Range Test. *Biometry*, 11: 1–48.
- [15] Tuhin J., Zinnat A. B. and Sayeeda S., 2007. Effect of Neem Oil on Some Pathogenic Bacteria. *Bangladesh Journal of Pharmacology*, 2 (2): 71–72.
- [16] Steel, R. G. D. and Torrie, J. H., 1984. Principles and procedures of statistics. Inten. student ed. Tokyo: McGraw Hill.
- [17] Etim N. N., Williams E. D., Akpabio U. and Offiong E. E. A., 2014. Haematological parameters and factors affecting their values. *Agricultural Science*, 2 (1): 37-47.
- [18] Isaac L. J., Abah G., Akpan B. and Ekaette I. U., 2013. Haematological properties of different breeds and sexes of rabbits. *Proceeding of the 18th Annual conference of Animal Science Associated of Nigeria*. 24-27.
- [19] Babatunde G. M., Fajimi A. O. and Oyejide A. O., 1992. Rubber seed oil versus palm oil in broiler chicken diets: effect on performance, nutrient digestibility, haematology and carcass characteristics. *Animal feed Science and Technology*, 35: 133-146.
- [20] Njidda A. A. and Isidahomen C. E., 2010. Hematology, blood chemistry and carcass characteristics of growth rabbits fed grasshopper meal as a substitute for fosh meal. *Pakistan Veterinary Journal*, 30 (1): 7-12.
- [21] Yasar N. F., Uylas M. U., Baspinar M. Sarsilmaz H., Ates E., Erkasap S. and Sahin A., 2016. Evaluating the use of heamatological parameters in staging hidradenitis suppurativa. *Wounds*, 28 (11): 87-91.
- [22] Vivian U., Oleforuh-Okoleh, Harriet M., Ndofor-Foleng, Salomon O., Olorunleke and Joseph O. U., 2015. Evaluation of growth performance, haematological and serum biochemical responses of broiler chickens to aqueous extract of ginger and garlic. *Journal of Agricultural Science*, 7 (4): 167-173.
- [23] Odoh L. I. and Lawrence L., 2015. Effects of varying levels of neem (*Azadirachta indica*) leaf meal in layer diets on the haematological and serological indices, and faecal bacteria counts of layers. *Journal of Natural Sciences Research*, 5 (4): 37-44.
- [24] Almosni-Le Sueur F., Dolz R., Natalia Majo N., 2011. Autopsie des volailles. Edt. Point Vétérinaire. Collection: Atlas Point Vétérinaire. 110p.
- [25] Divya K., Ankit K., Tripathi J. S. and Tiwari S. K., 2013. Evaluating anti-asthmatic effect of polyherbal ayurvedic drug bharangyadi on respiratory mechanics using matlab. *International Research Journal of Pharmacy*, 4 (2): 167-169.
- [26] Abalaka M., Oyewole O. A. and Kolawole A. R., 2012. Antibacterial Activities of *Azadirachta indica* against Some Bacterial Pathogens. *Advances in Life Sciences*, 2 (2): 5-8.